# CZECH MYCOLOGY

Publication of the Czech Scientific Society for Mycology

Volume 51

January 1999

Number 1

# Teleomorph-anamorph connections in Ascomycetes. 1. Cylindrotrichum and Cacumisporium anamorphs of Chaetosphaeria

# Martina Réblová<sup>1</sup> and Walter Gams<sup>2</sup>

<sup>1</sup>Institute of Botany, Academy of Sciences, 252 43 Průhonice, Czech Republic <sup>2</sup>Centraalbureau voor Schimmelcultures, P. O. Box 273, 3740 AG Baarn, The Netherlands

Réblová M. and Gams W. (1999): Teleomorph-anamorph connections in Ascomycetes 1. Cylindrotrichum and Cacumisporium anamorphs of Chaetosphaeria. – Czech Mycol. 51: 1–40

The teleomorph-anamorph connections of Chaetosphaeria are discussed. On the basis of the revision of the type and other herbarium material, Zignoëlla crustacea, Lasiosphacria britzelmayri subsp. fennica, Sphaeria decastyla and Melanomma macrosporum proved to be members of Chaetosphaeria. The new combinations Chaetosphaeria crustacea, Chaet. fennica and Chaet. decastyla, are proposed, Melanomma macrosporum is included in the synonymy of the latter. Chaetosphaeria crustacea has an anamorph intermediate between Chloridium and Cylindrotrichum, Chaetosphaeria fennica has a Chloridium-like anamorph, and Chaetosphaeria decastyla has Cacumisporium capitulatum as its anamorph. Two new species, Chaetosphaeria tulasneorum associated with the anamorph Cylindrotrichum oligospermum and Chaetosphaeria acutata associated with a Cylindrotrichum anamorph, are described. The teleomorph-anamorph connections of all accepted Chaetosphaeria species, except for Chaet. fennica, were confirmed by culture studies. Chaetosphaeria abietis and Chaet. fusiformis are also considered, and the connection with the Chloridium cylindrosporum anamorph of the latter is confirmed by culture studies for the first time. The genus Cylindrotrichum and its relationship to Chloridium and other related genera is discussed. A key to the species of Chaetosphaeria with Cylindrotrichum and Cacumisporium anamorphs is provided.

Key words: Ascomycetes, Chaetosphaeria, culture studies, wood-inhabiting fungi, systematics.

Réblová M. and Gams W. (1999): Vztahy teleomorfa-anamorfa u askomycetů 1. Cylindrotrichum a Cacumisporium, anamorfní rody rodu Chaetosphaeria. – Czech Mycol. 51: 1–40

Jsou diskutovány teleomorfní-anamorfní vztahy rodu Chaetosphaeria. Na základě revize typového a dalšího herbářového materiálu jsou druhy Zignoëlla crustacea, Lasiosphaeria britzelmayri subsp. fennica, Sphaeria decastyla a Melanomma macrosporum umístěny do rodu Chaetosphaeria. Jsou navrženy tři nové kombinace, a to Chaetosphaeria crustacea, Chaet. fennica a Chaet. decastyla. Melanomma macrosporum je zařazena do synonymiky druhu Chaet. decastyla. Rodové zařazení anamorfy druhů Chaetosphaeria crustacea a Chaet. fennica je na pomezí rodů Cylindrotrichum a Chloridium. Anamorfou Chaetosphaeria decastyla je Cacumisporium capitulatum. Jsou popsány dva nové druhy, Chaetosphaeria tulasneorum s anamorfou

Cylindrotrichum oligospermum a Chaetosphaeria acutata s anamorfou Cylindrotrichum sp. Teleomorfni-anamorfni vztahy všech výše uvedených druhů rodu Chaetosphaeria byly ověřeny kultivačními studiemi. Jsou zahrnuty také druhy Chaetosphaeria abietis a Chaet. fusiformis. Spojení Chaet. fusiformis s anamorfou Chloridium cylindrosporum je poprvé potvrzeno kultivačními studiemi. Jsou diskutovány vztahy rodu Cylindrotrichum s rodem Chloridium a dalšímí příbuznými rody. Je vytvořen klíč pro určování druhů Chaetosphaeria s anamorfami Cylindrotrichum a Cacumisporium.

#### INTRODUCTION

The genus *Chaetosphaeria* Tul. et C. Tul. (Tulasne and Tulasne 1863) accommodates lignicolous, saprobic fungi with superficial, dark, glabrous or sometimes setose, non-collapsing perithecia, the perithecial wall is brittle, consisting of thinwalled, opaque, brick-like cells or cells that form a network (*textura epidermoidea*); paraphyses and periphyses are persistent, asci have a distinct non-amyloid, refractive apical annulus and ascospores are transversely 1- to multi-septate, nonfragmenting and hyaline. Associated anamorphs have pigmented conidiophores and phialidic conidiogenesis, and are classified in several genera of hyphomycetes.

Among the synonyms of *Chaetosphaeria*, Zignoëlla Sacc. was characterized by hyaline, phragmosporous ascospores (Saccardo 1878), a criterion not recognized here as having generic relevance. Zignoëlla is a broadly conceived genus (Saccardo 1878, 1883) whose lectotype species, Zignoëlla pulviscula (Currey) Sacc., was selected only by Clements and Shear (1931) and was recombined by Booth (1957) in *Chaetosphaeria*.

Barr (1990) and Eriksson and Hawksworth (1993) placed Chaetosphaeria in the broadly perceived Lasiosphaeriaceae Nannf. Recently, Chaetosphaeria and six other related genera, viz. Ascocodinaea Samuels et al., Melanochaeta E. Müll. et al., Melanopsammella Höhnel, Porosphaerella E. Müll. et Samuels, Porosphaerellopsis Samuels et E. Müll., and Striatosphaeria Samuels et E. Müll., were transferred to a new family, the Chaetosphaeriaceae Réblová et al., in the order Sordariales Chadef. ex D. Hawksw. et O. E. Erikss. (Réblová et al. 1999). The Chaetosphaeriaceae are separated from the more narrowly delimited Lasiosphaeriaceae Nannf. and its core genera, viz. Lasiosphaeria Ces. et De Not., Bombardia (Fr.) P. Karst., Eosphaeria Höhnel, Cercophora Fuckel, and Apiosordaria Arx et W. Gams, on the basis of characters of perithecium, perithecial wall, asci and ascospores. Réblová et al. (1999) recognized twentytwo species of *Chaetosphaeria* with associated dematiaceous hyphomycetous anamorphs in nine genera, viz. Catenularia Grove, Cylindrotrichum Bonord., Chalara (Corda) Rabenh., Chloridium Link: Fr., Custingophora Stolk et al., Dictyochaeta Speg., Menispora Pers.: Fr., Phialophora Medlar, and Zanclospora S. Hughes et B. Kendrick (Saccardo 1883; Booth 1957, 1958; Müller and von

Arx 1962; Hughes and Kendrick 1968; Gams and Holubová-Jechová 1976; Barr and Crane 1979; Holubová-Jechová 1973, 1982, 1984; Barr 1993; Constantinescu et al. 1995; Réblová 1998 a, b).

The genus Cylindrotrichum was discussed by a number of authors (e.g. Hughes 1951; Gams and Holubová-Jechová 1976; Morgan-Jones 1977, 1980; Kendrick 1980; DiCosmo et al. 1983; Cabello and Arambarri 1988; Holubová-Jechová 1990), but no clear and final generic concept is available. The genus comprises species with erect, unbranched conidiophores, terminal conidiogenous cells with multiple or single conidiogenous loci produced by sympodial or percurrent proliferation and hyaline, cylindrical to long ellipsoidal, 1-3-septate conidia. DiCosmo et al. (1983) placed species of the genus Cylindrotrichum into two new genera, Kylindria DiCosmo et al. and Xenokylindria DiCosmo et al., and transferred one species formerly placed in Cylindrotrichum to Uncigera Sacc., two to Dictyochaeta Speg. and five to Chaetopsis Greville emend. DiCosmo et al., including the type species, Cylindrotrichum oligospermum (Corda) Bonord. These authors emphasized especially the shape of the conidia and the conidiogenous cells, the distinction between mono- and polyphialides being a main differentiating character. They preferred to distinguish several closely related anamorph genera of Chaetosphaeria, such as Chaetopsis, Dictyochaeta, Kylindria and Xenokylindria, based on details of conidiogenesis, though they share a number of morphological, developmental and ecological characteristics (DiCosmo et al. 1983).

Cabello and Arambarri (1988) considered the concept proposed by DiCosmo et al. (1983) incorrect. They redefined the generic concept of *Cylindrotrichum* and accepted eleven species. The genera *Kylindria* and *Xenokylindria* were reduced to synonyms of *Cylindrotrichum*. According to Cabello and Arambarri (1988), *Cylindrotrichum* accommodates species formerly placed in three closely related genera, viz. *Kylindria, Xenokylindria* and *Chaetopsis*, that are associated with one teleomorph genus, *Chaetosphaeria*. Although they did not consider teleomorphanamorph connections of any of the species accepted in *Cylindrotrichum*, their generic concept of *Cylindrotrichum* justifies the close relationship of the *Chaetosphaeria* anamorphs better than the system proposed by DiCosmo et al. (1983).

In the present paper we describe five species of *Chaetosphaeria*, four of them having *Cylindrotrichum* anamorphs and one a *Cacumisporium* Preuss anamorph. Three species were previously known as *Zignoëlla crustacea* Sacc. (Saccardo 1883), *Lasiosphaeria britzelmayri* Sacc. subsp. *fennica* P. Karst. (Karsten 1887) and *Sphaeria decastyla* Cooke (Cooke 1878) [= *Melanomma macrosporum* Sacc. (Saccardo 1875)]. The respective type specimens were examined. On the basis of perithecial anatomy, ascal, ascospore and hamathecium anatomy, and conidiogenesis of the anamorphs, these three species belong to *Chaetosphaeria*. They are recombined as *Chaetosphaeria crustacea*, *Chaet. fennica* and *Chaet. decastyla*. *Melanomma macrosporum* is included in the synonymy of the latter. These species,

#### Czech mycol. 51 (1), 1999

except for *Chaet. fennica*, were recollected on strongly decayed wood of coniferous and deciduous trees in Europe (France, Czech Republic, Ukraine). They are redescribed and illustrated along with notes on previous descriptions and illustrations. The anamorph of *Chaet. crustacea* belongs to *Cylindrotrichum*, though it shows great similarity with *Chloridium*. The anamorph of *Chaet. fennica* is *Chloridium*-like. The anamorph of *Chaet. decastyla* is *Cacumisporium capitulatum* (Corda) S. Hughes. On the basis of conidiogenesis and the structure and function of the phialides, *Cac. capitulatum* may also be included in *Cylindrotrichum*, but we refrain at the moment from making this combination before the anamorphgeneric delimitations are further analysed. With the exception of *Chaet. fennica*, conidiophores of all these species were observed on a natural substratum and in living cultures.

Several collections of another fungus associated with a *Cylindrotrichum* anamorph were made on decayed wood of deciduous trees in the Czech Republic, France and Ukraine. It has fusiform, hyaline, at maturity 3-septate ascospores, persistent paraphyses and periphyses, unitunicate asci, black glabrous perithecia with a fragile wall and a dematiaceous hyphomycete anamorph with phialidic conidiogenesis. The anamorph was also obtained in culture. This fungus belongs to *Chaetosphaeria* and is described here as *Chaet. acutata*.

A single collection of the fifth species was made on a dead branch of Sambucus nigra in the Czech Republic. The anamorph that was obtained in culture represents Cylindrotrichum oligospermum (Corda) Bonord., the type of the generic name Cylindrotrichum. On the basis of the black, glabrous perithecia with brittle wall, persistent paraphyses and periphyses, ascospores that are hyaline, 1–3-septate and fusiform, and the anamorph, this recent collection represents an undescribed species of Chaetosphaeria, which is described here as Chaet. tulasncorum.

Chactosphaeria abietis (Höhnel) W. Gams et Holubová-Jechová and Chaetosphaeria fusiformis W. Gams et Holubová-Jechová (Gams and Holubová-Jechová 1976, 1981) are discussed. A key to the species of Chaetosphaeria with Cylindrotrichum and Cacumisporium anamorphs is provided.

### MATERIAL AND METHODS

Dry herbarium specimens were rehydrated in 3% (aq.) KOH and subsequently studied in water, Congo Red (aq.) and Melzer's reagent. The abbreviations of herbaria and institutes that kindly lent the material are cited in accordance with the Index Herbariorum (Holmgren et al. 1990).

In the lists of material examined M. R. is the abbreviation for M. Réblová.

Single ascospores of *Chaetosphaeria crustacea*, *Chaet. acutata*, *Chaet. fusiformis*, *Chaet. decastyla* and *Chaet. tulasneorum* were isolated with the aid of a single-spore isolator on cornmeal agar (CMA, Difco). Colonies were grown

on CMA, malt extract agar (MEA), oatmeal agar (OA) and potato-carrot agar (PCA), colony characters were taken from PCA cultures grown for 10 days at 25 °C in darkness and 10 days at 25 °C in cool white fluorescent light. The cultures are maintained in the Institute of Botany, Academy of Sciences at Průhonice and the Centraalbureau voor Schimmelcultures (CBS) at Baarn, the Netherlands.

# TAXONOMIC PART

Key to the species of *Chaetosphaeria* with *Cylindrotrichum* and *Cacumisporium* anamorphs

1.	Occurring on decayed wood and bark of angiosperms2
1.	Occurring on decayed wood and bark of gymnosperms
	2. Ascospores at maturity 3-septate 3
3.	Ascospores short fusiform, not exceeding 21 $\mu$ m; anamorph Cylindrotrichum oligospermum
3.	<ul> <li>Ascospores elongate-fusiform, longer than 30 μm</li></ul>
5.	Ascospores cylindrical-fusiform, 3–5-septate, perithecia setose; anamorph Cylin- drotrichum-Chloridium-like
5.	<ul> <li>Ascospores cylindrical-fusiform, at maturity 3-septate, perithecia glabrous6</li> <li>6. Ascospores asymmetric, tapering towards one end and rounded at the other end; anamorph <i>Chloridium cylindrosporum</i>5. <i>Chaetosphaeria fusiformis</i></li> <li>6. Ascospores symmetric with rounded ends; anamorph <i>Cylindrotrichum zig- noëllae</i>1. <i>Chaetosphaeria abietis</i></li> </ul>

 Chaetosphaeria abietis (Höhnel) W. Gams et Hol.-Jech., Stud. Mycol. 13: 53, 1976.
 Fig. 1a-f.

≡ Zignoëlla abietis Höhnel, in Rehm, Ann. Mycol. 5: 469, 1907; more elaborated in Höhnel, Sitzungsber. K. Akad. Wiss. Wien, Math.-naturw. Kl., Abt. 1, 118: 332, 1909.

Anamorph. Cylindrotrichum zignoëllae (Höhnel) W. Gams et Hol.-Jech., Stud. Mycol. 13: 53, 1976. Czech mycol. 51 (1), 1999

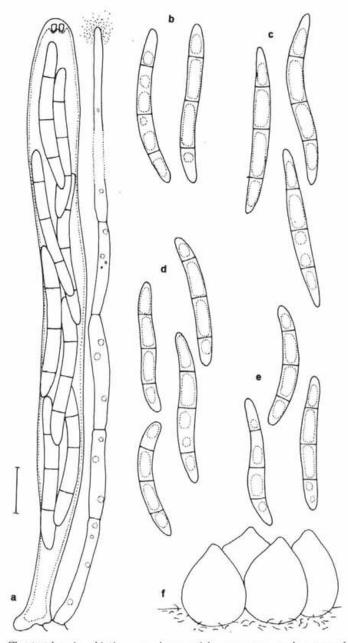


Fig. 1a–f. Chaetosphaeria abietis. – a. Ascus with ascospores and a paraphysis. – b–e. Ascospores. – f. Habit sketch of a group of perithecia. – a, b, f from Herb. M. R. 1058/97; c from Herb. M. R. 957/97; d from Herb. M. R. 969/97; e from the type K 49555. – Scale bar: a-e = 10  $\mu$ m.

- ≡ Acrotheca zignoëllae Höhnel, Sitzungsber. K. Akad. Wiss. Wien, Math.naturw. Abt. 1, Kl. 118: 332, 1909.
- $\equiv$  Kylindria zignoëllae (Höhnel) DiCosmo et al., Mycologia 75: 971, 1983.
- = Cylindrotrichum oblongisporum G. Morgan-Jones, Mycotaxon 5: 487, 1977.

Material examined. 1) Type material. Rehm Ascom. exsicc. No. 1740 (AUS-TRIA. Dürrwien, Wiener Wald, on inner side of decayed bark, F. von Höhnel, July 1907, K 49555 – lectotype of Zignoëlla abietis).

2) Additional material. Czech Republic. Southern Bohemia, Šumava Mts., glacial cirque of the lake Čertovo jezero near Železná Ruda, on inner surface of bark of *Abies alba*, 28 Aug. 1997, M. R. (Herb. M. R. 1027/97); ibid., glacial cirque of the lake Černé jezero near Železná Ruda, on bark of *Abies alba*, 27 Aug. 1997, M. R. (Herb. M. R. 1058/97). – France. Pyrenees, Quérigut, Laurenti Lake, on bark of *Abies alba*, 17 July 1997, M. R. (Herb. M. R. 969/97, 973/97). – Ukraine. Eastern Carpathian Mts., Kvasi near Rachiv, on left bank of the river Tisa, on inner side of bark of *Abies alba*, 28 June 1997, M. R. (Herb. M. R. 957/97).

Descriptions and illustrations. Rehm (1907: 469); Höhnel (1909: 332); Gams and Holubová-Jechová (1976: 53, Fig. 26 a, b).

Habitat. Saprobe on decayed wood and inner surface of bark of conifers. Known host. *Abies alba*.

Known distribution. Europe: Austria, Czech Republic, France, Ukraine.

Note. The tips of paraphyses were seen enclosed in a small gelatinous cap that was well visible in phase contrast. This character was not seen in any other *Chaetosphaeria* species.

Chactosphaeria abietis has not yet been grown in culture. The associated anamorph Cylindrotrichum zignoëllae was regularly observed growing around the perithecia (Gams and Holubová-Jechová 1976; Réblová, unpublished observations). Hawksworth and Minter (1980) reported the anamorph from dead herbaceous material of Filipendula ulmaria to be associated with perithecia of a Chactosphaeria-like fungus; its ascospores were described as differing from Chaet. abietis, being 1–3-septate, shortly fusiform, 13–17 × 3–4  $\mu$ m. Hawksworth and Minter (1980) concluded that Cylindrotrichum zignoëllae sensu Gams and Holubová-Jechová (1976) was circumscribed too broadly and may accommodate two taxa belonging to distinct teleomorphs. The perithecia found on Filipendula ulmaria could not be investigated further, for most of them were immature. No attempt was made to cultivate the specimen (Hawksworth and Minter 1980).

*Chaetosphaeria abietis* occurs rarely. All recent collections were made in regions with natural stands of *Abies alba*. Ascospore germination was not observed.

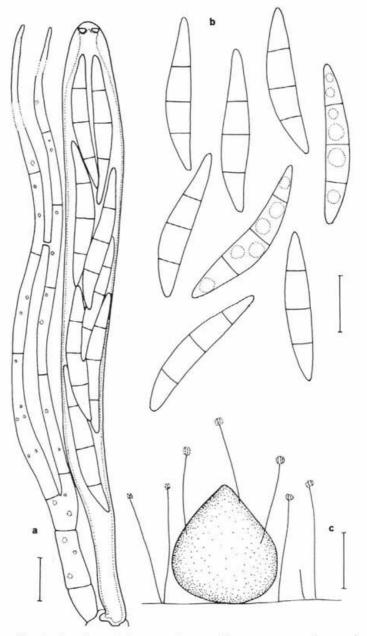


Fig. 2a–c. Chaetosphaeria acutata. – a. Ascus with ascospores and paraphyses. – b. Ascospores. – c. Habit sketch of the perithecium and conidiophores. – a–c from the type PRM 842979. – Scale bars: a, b = 10  $\mu$ m; c = 100  $\mu$ m.

Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

# 2. Chaetosphaeria acutata Réblová et W. Gams, sp. nov.

Figs. 2, 3.

Perithecia superficialia, gregaria vel solitaria, subglobosa, 170–220  $\mu$ m alta, 170–200  $\mu$ m diam, papillata, ostiolata, fusca vel quasi atra, lucida, glabra vel conidiophoris obtecta, modice asperulata. Canalis ostiolaris periphysatus. Paries perithecii fragilis, ad latus 26–33  $\mu$ m crassus, bistratosus: stratum exterius e cellulis tenuitunicatis, fuscis, lateriformibus constans, interius e cellulis tenuitunicatis, compressis, subhyalinis. Paraphyses copiosae, intervallis 11–21  $\mu$ m septatae, ad septa non constrictae, ramosae, anastomosantes, hyalinae, 3–4  $\mu$ m latae in parte inferiore, ad 1.5–2  $\mu$ m angustatae, ultra ascorum apices protrudentes. Asci 8-spori, cylindrici vel clavati, (95–)121–147 × (8.5–)9.5–10.5(–11.5)  $\mu$ m, breviter stipitati, apex anulo I-, refringente, 3.0–3.5  $\mu$ m lato et 1.0–1.2  $\mu$ m alto praeditus, 8-spori. Ascosporae fusiformes, utrinque fere acutatae et minime rotundatae, (28–)30.5–38(–44) × 3–4(–5)  $\mu$ m, 3-septatae, non constrictae, leves, hyalinae, oblique 1–2(–3)-seriatae in asco, tubis polaribus germinantes.

Anamorphosis Cylindrotrichum sp.: Conidiophoris simplicibus, fuscis, sursum pallidioribus, 125–190 × (4–)5–6.5  $\mu$ m, sursum ad 3–4  $\mu$ m angustatis, monophialidicis, saepe semel percurrenter proliferentibus. Phialides supra collare 4–10.5  $\mu$ m extendentes, quinquies ad dodecies annellatae. Conidia cylindrica vel clavata, recta vel modice curvata, ad basim truncata, ad apicem rotundata, 1-septata, haud constricta, levia, hyalina, (11–)12.5–15.5(–16.5) × 4–5  $\mu$ m.

Holotypus. Bohemia meridionalis, montes Šumava, Železná Ruda, ad lignum putridum *Fagi sylvaticae*, 28 Aug. 1997, leg. M. R. 994/97 (PRM 842979).

Etymology. Lat. acutatus = pointed, referring to the tapering ends of the ascospores.

Anamorph. Cylindrotrichum sp. (described here). Fig. 3a-d.

Teleomorph. Perithecia superficial, gregarious to solitary, subglobose, 170– -220  $\mu$ m high and 170–200  $\mu$ m diam, papillate, ostiolate, dark brown to nearly black, glistening, glabrous or covered with conidiophores of the anamorph, slightly roughened. Perithecial wall brittle, lateral wall 26–33  $\mu$ m thick, consisting of two layers; an outer layer of thin-walled, dark brown, opaque brick-like cells, and an inner layer of thin-walled, compressed, subhyaline cells. Ostiolar canal periphysate. Paraphyses copious, septate at 11–21  $\mu$ m intervals, non-constricted at the septa, branching, anastomosing, 3–4  $\mu$ m wide in the lower part, tapering to 1.5–2  $\mu$ m, protruding beyond the tips of the asci. Asci cylindrical-clavate, (95–)121–147 × (8.5–)9.5–10.5(–11.5)  $\mu$ m, shortly stipitate, ascal apex with a J-, refractive, 3–3.5  $\mu$ m wide and 1–1.2  $\mu$ m deep apical annulus. Ascospores fusiform, narrowly rounded at the ends, (28–)30.5–38(–44) × 3–4(–5)  $\mu$ m, 3-septate, not

CZECH MYCOL. 51 (1), 1999

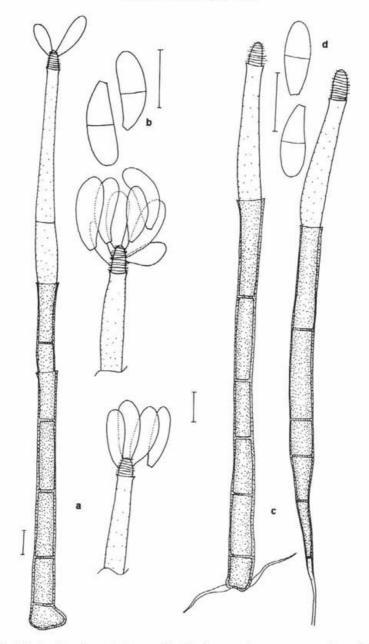


Fig. 3a–d. Chaetosphaeria acutata. – a. Conidiophore and sporogenous apices of the conidiophores, from nature. – b. Conidia, from nature. – c. Conidiophores, from PCA culture. – d. Conidia, from PCA culture. – a–d from CBS 101315, PRM 842979. – Scale bar: a–d = 10  $\mu$ m.

constricted, smooth, hyaline, obliquely 1-2(-3)-seriate, partially overlapping in the ascus, germinating by germ tubes at the ends.

Anamorph. Colonies hairy, dark, conidiophores arising from the substratum or covering the perithecia. Setae absent. Conidiophores macronematous, mononematous, solitary, erect, unbranched, cylindrical, 125–190  $\mu$ m long and (4–)5–6.5  $\mu$ m wide in the middle, tapering to 3–4  $\mu$ m, straight or slightly flexuous, dark brown, paler towards the apex, smooth, often with 1 percurrent proliferation. Phialides terminal, cylindrical, the percurrently proliferating part above the collarette 4–10.5  $\mu$ m long and 3.5–5  $\mu$ m wide, with 5–12 densely annellate proliferations. Primary collarettes hyaline, 4.5–5.5(–6)  $\mu$ m wide and 1  $\mu$ m deep. Conidia (11–)12.5–15.5(–16.5) × 4–5  $\mu$ m, cylindrical to clavate, straight or slightly curved, tapering and truncate at the proximal end, rounded at the distal end, 1-septate, not constricted, smooth-walled, hyaline.

Characteristics in culture. Colonies on PCA slow-growing, reaching 3–4 mm diam in 10 days at 25 °C in darkness, when grown for another 10 days at 25 °C in cool white fluorescent light reaching 7–8 mm diam; felty, greyish brown, aerial mycelium developed, margins fimbriate, no conidiation; reverse dark brown to nearly black. In 20 days: CMA: 4–4.5 mm diam, MEA: 6–7 mm diam; OA: 3.5–4 mm diam; CMA, MEA: felty, greyish brown, aerial mycelium well-developed, margins fimbriate, no conidiation. Colonies having a moist appearance, midbrown, aerial mycelium scant, no conidiation. Colonies sporulated only on PCA, OA and CMA in 2–3-month-old slant cultures preserved at 18 °C under cool white fluorescent light. Mycelium superficial or immersed; hyphae branched, septate, subhyaline, smooth, 2.5–3  $\mu$ m wide. Setae absent. Conidiophores as on the natural substratum 84–150(–200)  $\mu$ m long and 5–6  $\mu$ m wide in the middle, tapering to 3–4.5  $\mu$ m. Phialides with hyaline primary collarette 4–6  $\mu$ m wide and 1–1.5(–2)  $\mu$ m deep. Conidia 10.5–14.5  $\times$  3–4.5  $\mu$ m.

Material examined. 1) Type material. Czech Republic. Southern Bohemia, Šumava Mts., glacial cirque of the lake Čertovo jezero near Železná Ruda, on wood of *Fagus sylvatica*, 28 Aug. 1997, M. R. 994/97 (PRM 842979 – holotype of *Chaetosphaeria acutata*)

2) Additional material examined. Czech Republic. Southern Bohemia, Šumava Mts., glacial cirque of the lake Černé jezero near Železná Ruda, on wood of *Fagus sylvatica*, 23 Oct. 1996, M. R. (Herb. M. R. 906/96); ibid., 27 Aug. 1997, M. R. (Herb. M. R. 1043/97). – France. Central Pyrenees, Bagnères de Luchon, Lys valley, on wood of *Fagus sylvatica*, 13 July 1997, M. R. (Herb. M. R. 974/97). – Ukraine, Eastern Carpathian Mts., Kvasi near Rachiv, on left bank of the river Tisa, on wood of *Corylus avellana*, 26 June 1997, M. R. (Herb. M. R. 948/97).

#### Czech mycol. 51 (1), 1999

Cultures. CBS 101311 (Herb. M. R. 948/97); CBS 101312 (Herb. M. R. 974/97); CBS 101315 (PRM 842979).

Habitat. Saprobe on decayed wood of deciduous trees.

Known hosts. Corylus avellana, Fagus sylvatica.

Known distribution. Europe: Czech Republic, France, Ukraine.

Note. The species was initially identified as Lasiosphaeria britzelmayri Sacc. subsp. fennica P. Karst. (Karsten 1887). The redescription of this taxon by Podlahová (1974) seemed to match our fungus perfectly. But the type specimen in H showed that ascospores clearly differ in shape and size. They are rather longer  $[(34.5-)36.5-42(-43) \times (3.5-)4(-4.5) \mu m]$ , elongate-fusiform and tapering strongly towards the ends immediately after the middle septum. The asci are of a size  $[(126-)133-152(-168) \times (8.5-)9-10.5 \mu m]$  comparable to that of *Chaetosphaeria* acutata. The anamorph associated with perithecia on the type material of Lasiosphaeria britzelmayri subsp. fennica is *Chloridium*-like and entirely different from the *Cylindrotrichum* anamorph of *Chaet. acutata*. The conidia are mid-brown and non-septate. Lasiosphaeria britzelmayri Sacc. subsp. fennica is therefore another species of *Chaetosphaeria* to be included in the present paper.

Chaetosphaeria acutata clearly differs from other species of the genus by its typically long fusiform, at maturity 3-septate ascospores, the *Cylindrotrichum* anamorph and its exclusive occurrence on decayed wood of deciduous trees. In areas with near-natural and natural stands dominated by *Fagus sylvatica* and *Abies alba*, where the fungus occurs, it is not uncommon.

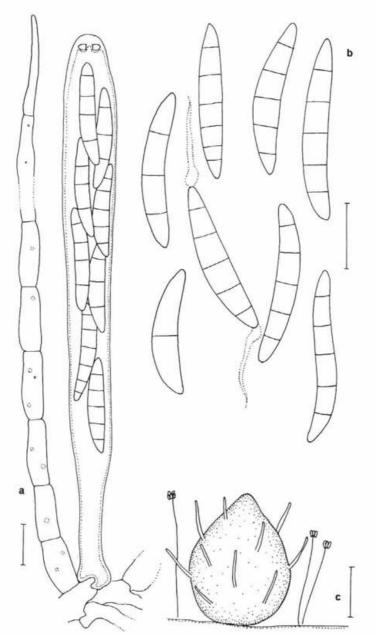
Conidia of the *Cylindrotrichum* anamorph of *Chaet. acutata* are formed successively from the sporogenous apex of the proliferating conidiogenous cells. *Cacumisporium capitulatum* has similarly proliferating conidiogenous cells but differs in having at maturity 4-celled, bicolorous and larger conidia.

### 3. Chaetosphaeria crustacea (Sacc.) Réblová et W. Gams, comb. nov. Figs. 4-6.

- ≡ Zignoëlla crustacea Sacc., Syll. Fung. 2: 220, 1883. basionym.
- ≡ Sphaeria crustacea P. Karst., Fungi fenn. exs. 865, 1869; more elaborated in Bidr. Känn. Finl. Nat. Folk. 23: 95, 1873 [non Sphaeria crustacea Sow., Col. Fig. Engl. Fung. 1: Tab. 372, Fig. 3, Pl. 372, 1803 = Hypoxylon multiforme (Fr.: Fr.) Fr.].

Anamorph. Intermediate between *Cylindrotrichum* and *Chloridium* (described here). Fig. 6a–f.

Teleomorph. Perithecia superficial, solitary to densely gregarious, subglobose to globose, 130–250  $\mu$ m high and 120–220  $\mu$ m diam, papillate, ostiolate, papilla



Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

Fig. 4a–c. Chaetosphaeria crustacea. – a. Ascus with ascospores and a paraphysis. – b. Ascospores. – c. Habit sketch of perithecium and conidiophores. – a–c from Herb. M. R. 996/97. – Scale bars: a, b = 10  $\mu$ m; c = 100  $\mu$ m.

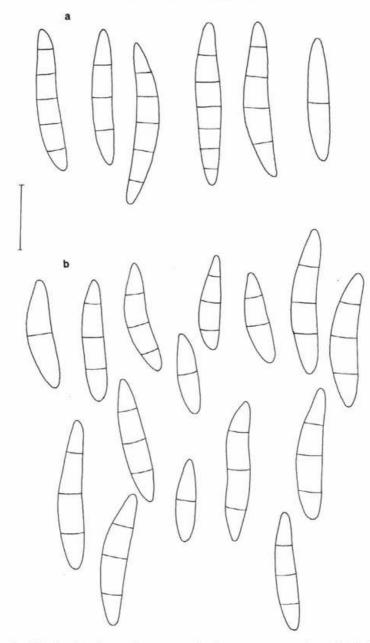


Fig. 5a, b. Chaetosphaeria crustacea. – a, b. Ascospores. – a from H; b from Herb. M. R. 1169/97. – Scale bar: a, b = 10  $\mu m.$ 

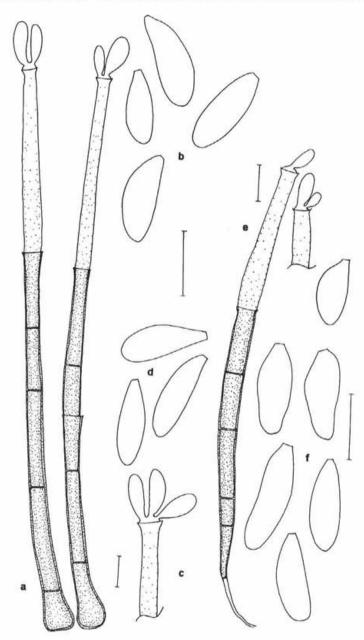


Fig. 6a–f. Chaetosphaeria crustacea. – a. Conidiophores, from nature. – b. Conidia, from nature. – c. Sporogenous apex of the conidiophore, from PCA culture. – d, f. Conidia, from PCA culture. – e. Conidiophore, from PCA culture. – a, b, e, f from Herb. M. R. 996/97; c, d from CBS 101321, Herb. M. R. 1169/97; e, f from CBS 101316. – Scale bar:  $a-f = 10 \ \mu m$ .

perforated by a rounded pore in old perithecia, black, glistening, setose, slightly roughened. Setae scattered over the perithecia, erect, dark brown, septate, obtuse, 30–130  $\mu$ m long and 3–3.5  $\mu$ m wide in the middle. Perithecial wall brittle, lateral wall 15–22  $\mu$ m thick, consisting of two layers; an outer layer of thin-walled, opaque, brick-like cells, and an inner layer of thin-walled, subhyaline, compressed cells. Ostiolar canal periphysate. Paraphyses copious, septate at 18–21  $\mu$ m intervals, slightly constricted at the septa, branching, anastomosing, 4–5  $\mu$ m wide in the lower part, tapering to 2  $\mu$ m, protruding beyond the tips of the asci. Asci cylindrical-clavate, 75–105(–115) × (8.5–) 9.5–10.5  $\mu$ m, shortly stipitate, 8-spored, ascal apex with a J-, 2.5  $\mu$ m wide and 1.5  $\mu$ m deep apical annulus. Ascospores cylindrical-fusiform, straight or curved, (15.5–)19–26(–28) × 3–3.5(–4)  $\mu$ m, predominantly 3-septate, with a delayed formation of the 2–3 additional septa, not constricted, smooth, hyaline, 2–3-seriate, partially overlapping in the ascus, germinating by a germ tube at one or both ends; no tendency of fragmentation.

Anamorph. Colonies hairy, dark, conidiophores arising from decayed wood and also covering the perithecia. Setae absent. Conidiophores macronematous, mononematous, solitary, erect, unbranched, 170–210  $\mu$ m long and 4.5–5.5  $\mu$ m wide above the base, tapering to 3–3.5  $\mu$ m below the collarette, cylindrical, straight or slightly flexuous, septate, often with 1(–2) distant percurrent proliferation, dark brown, paler towards the apex, smooth-walled. Phialides terminal, cylindrical, 23–32 × 3.5–4.5  $\mu$ m. Collarettes hyaline, 4–5  $\mu$ m wide and 1.5–2.5  $\mu$ m deep. Conidia arising in a sympodial manner on a central dome inside the collarette like in *Chloridium virescens*, cylindrical-clavate, (11.5–)14.5–17(–21) × 3–4.5  $\mu$ m, straight or curved, rounded at the tip, tapering and truncate at the base, 1-celled, smooth, hyaline.

Characteristics in culture. Colonies on PCA slow-growing, reaching 4–5 mm diam in 10 days at 25 °C in darkness, when grown for another 10 days at 25 °C in cool white fluorescent light reaching 10–11 mm diam; velvety, centre dark greyish brown due to hyphae of aerial mycelium and conidiophores, dark brown to nearly black at the fimbriate margin, forming a conspicuous dark, sterile zone of submerged mycelium; conidial production copious in 10 days; conidiophores arising in the centre; conidial masses globose, whitish; reverse dark grey; no pigments released. In 20 days: CMA: 9–10 mm diam; MEA 12 mm diam; OA 9–12 mm diam; CMA, MEA, OA: velvety, dark grey, with copious conidial production in 10 days on CMA, OA; on MEA after 20 days. The black sterile zone formed by substrate mycelium at the margins is most pronounced on CMA, less on PCA and OA and is not developed on MEA. Mycelium superficial or immersed; hyphae branched, septate, subhyaline, smooth, 2–3  $\mu$ m wide. Setae absent. Conidiophores as on the natural substratum, 70–160  $\mu$ m

Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

long, 4–5  $\mu$ m wide in the middle, tapering to 3–3.5  $\mu$ m below the collarette. Phialides 23–44 × 3.5–4.5  $\mu$ m; collarettes 4–5  $\mu$ m wide and 1.5–2.5(–3)  $\mu$ m deep. Conidia (11–)12.5–16.5(–20) × 3–4(–5)  $\mu$ m. Chlamydospores absent.

Material examined. 1)Type material. Finland. Ostrobottnia, Vaasa, on *Pinus sylvestris*, P. A. Karsten, Fung. Fenn. exsicc. 865, (H – lectotype of Zignoëlla crustacea).

2) Additional material. Czech Republic. Southern Bohemia, glacial cirque of the lake Černé jezero near Železná Ruda, on wood of Abies alba, 7 Nov. 1997, K. Prášil (Herb. M. R. 1169/97). – Ukraine. Eastern Carpathian Mts., Kvasi near Rachiv, on left bank of the river Tisa, on wood of Picea abies, 26 June 1997, M. R. (Herb. M. R. 996/97).

Cultures. CBS 101316 (Herb. M. R. 996/97); CBS 101321 (Herb. M. R. 1169/97).

Descriptions and illustrations. Karsten (1873: 95); Saccardo (1883: 220).

Habitat. Saprobe on decayed wood and bark of conifers.

Known hosts. Abies alba, Picea abies, Pinus sylvestris.

Known distribution. Europe: Czech Republic, Finland, Ukraine.

Note. The type collection of Zignoëlla crustacea made by Karsten in Finland (Karsten 1873) contains densely setose perithecia seated on wood of Pinus sylvestris, and 3–5 transversely septate, hyaline ascospores  $(22-24.5 \times 3-3.5 \ \mu m)$ . No anamorph was observed. The perithecia from the recent collections are densely setose in a collection made in the Czech Republic and sparsely setose in a collection made in the Ukraine. In both collections the same Cylindrotrichum species was found growing on the substratum and was also obtained in the living culture. Traditionally, Chaetosphaeria only contains species with glabrous perithecia. They may, however, become setose when the setae and conidiophores of the associated anamorph arise both from the substratum and the perithecial surface. Chaetosphaeria crustacea is an example of a fungus with setose perithecia associated with a Cylindrotrichum-Chloridium-like anamorph. The recent finds represent the first records of this fungus since its description.

In the type material both 3-septate and 4–5-septate ascospores were present. Inside the asci only the 3-septate ascospores were seen. The 4–5-septate ascospores were released from the perithecium and seen attached to the perithecial surface and on the substratum. In the specimen from the Ukraine mainly 5-septate ascospores and in that from the Czech Republic only 3-septate ascospores were seen.

Chaetosphaeria crustacea is similar in ascospore shape and septation to Chaet. decastyla but differs in shorter ascospores and asci, setose perithecia, the Cylindrotrichum-Chloridium-like anamorph and the occurrence on decayed wood of conifers. Chaetosphaeria crustacea occurs rarely and is known from three localities in Europe only.

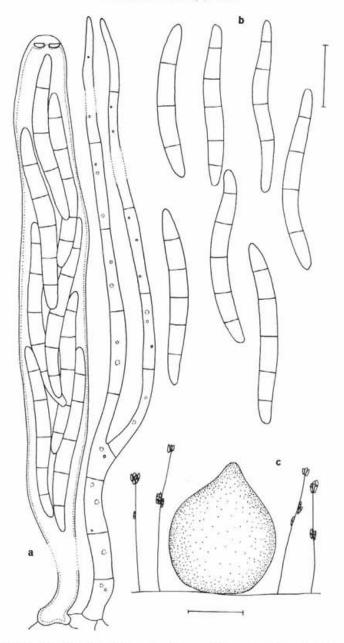


Fig. 7a–c. Chaetosphaeria decastyla. – a. Ascus with ascospores and paraphyses. – b. Ascospores. – Habit sketch of perithecium and conidiophores. – a–c from Herb. M. R. 985/97. – Scale bars: a, b = 10  $\mu m;$  c = 100  $\mu m.$ 

4. Chaetosphaeria decastyla (Cooke) Réblová et W. Gams, comb. nov.

Figs. 7-9.

- $\equiv$  Sphaeria decastyla Cooke, Grevillea 7: 52, 1878. basionym.
- $\equiv$  Acanthostigma decastylum (Cooke) Sacc., Syll. Fung. 2: 210, 1883.
- = Melanomma macrosporum Sacc., Hedwigia 14: 73, May 1875; see also Saccardo, Fungi ital. delin. 300, Feb. 1878; Michelia 1: 449, 15 Nov. 1878.
  - ≡ Zignoëlla macrospora (Sacc.) Sacc., Michelia 1: 346, 1 July 1878; Syll. Fung. 2: 221, 1883 [non Chaetosphaeria macrospora (Kawamura) Hara, J. Pl. Prot. Tokyo 16(2): 16, 1930].

Anamorph. Cacumisporium capitulatum (Corda) S. Hughes, Canad. J. Bot. 96: 743, 1958. Fig. 9a–d.

 $\equiv$  Helminthosporium capitulatum Corda, Ic. Fung. 2: 13, 1838.

= Cacumisporium tenebrosum Preuss, Linnaea 24: 130, 1851.

Teleomorph. Perithecia superficial, solitary or in groups of 3–5, subglobose to conical, 200–260  $\mu$ m high and 180–210  $\mu$ m diam, papillate, papilla perforated by a rounded pore in old perithecia, ostiolate, dark brown to black, glistening, glabrous or covered with conidiophores of the anamorph, slightly rugose. Perithecial wall brittle, 17–22  $\mu$ m thick, consisting of two layers; an outer layer of thin-walled, dark brown, opaque, polyhedral to brick-like cells, and an inner layer of thin-walled, compressed, hyaline cells. Ostiolar canal periphysate. Paraphyses copious, branching, septate at 18–42  $\mu$ m intervals, 3–3.5  $\mu$ m wide in the lower part, tapering to 1.5  $\mu$ m, protruding beyond the tips of the asci. Asci clavate-cylindrical, 68–90  $\mu$ m long in the pars sporifera × 10.5–11.5  $\mu$ m, stipe 30–54  $\mu$ m long, narrowly rounded at the tip, with a J-, refractive, 3–3.5  $\mu$ m wide and 1  $\mu$ m deep apical annulus. Ascospores cylindrical to cylindrical-fusiform or fusiform, occasionally tapering towards one end, (28–)30–42(-46) × 3–4  $\mu$ m, at maturity 5-septate, not constricted or very slightly constricted at the septa, smooth-walled, hyaline, 2–3-seriate, overlapping in the upper part of the ascus, germinating by germ tubes at the ends.

Anamorph. Colonies hairy, dark, conidiophores arising from the substratum or covering the perithecia. Setae absent. Conidiophores macronematous, mononematous, solitary, erect, unbranched, cylindrical, up to 200  $\mu$ m long, 6.5–7.5  $\mu$ m wide above the base, tapering to 5–6  $\mu$ m below the primary collarette, straight or slightly flexuous, septate, brown to pale brown, paler towards the apex, smooth, with 1–2 major percurrent proliferations. Phialides terminal, cylindrical, the proliferating part above the primary collarette 10–15  $\mu$ m long and 5–6  $\mu$ m wide, with 9–12 narrow annellate proliferations. Primary collarettes almost hyaline, 7–9.5  $\mu$ m wide and (1–)1.5–2  $\mu$ m deep. Conidia cylindrical, 15–20 × 5–6.5  $\mu$ m, straight or slightly curved, rounded at the tip, truncate at the base, 3-septate, not constricted, at

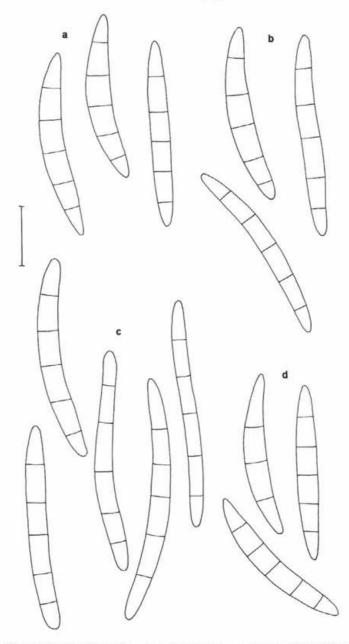


Fig. 8a–d. Chaetosphaeria decastyla. – a–d. Ascospores. – a–d from NY: a (USA, New Jersey, Newfield, on bark and wood of Magnolia sp.); b (USA, Georgia, Darien, on wood of Quercus sp., Ravenel 2420); c (Gloucester County, Newfield, 22 Jan. 1878); d (USA, Louisiana, on a decayed log of oak, 2 Jan. 1886, A. B. Langlois). – Scale bars: a–d = 10  $\mu$ m.

## Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

first hyaline, at maturity the two middle cells turning brown and the two end cells remaining hyaline, mature conidia often attached to the middle and lower part of the conidiophore.

Characteristics in culture. Colonies on PCA slow-growing, reaching 4–5 mm diam in 10 days at 25 °C in darkness, when grown for another 10 days at 25 °C in cool white fluorescent light reaching 8 mm diam; velvety, dark grey, aerial mycelium well-developed, margins fimbriate; conidial production scarce after 20 days; conidiophores scattered, conidial masses globose, hyaline; reverse greyish. Colonies sporulating copiously with conidiophores arranged in 2-3 concentric zones in 2 month-old slant cultures preserved at 18 °C under cool white fluorescent light. In 20 days: CMA: 7-8 mm diam; OA: 6-7 mm diam; CMA, OA: velvety, dark grey, conidial production scarce at the margins; MEA: 9-10 mm diam, velvety, greyish to pale brown, no conidial production. Mycelium superficial or immersed; hyphae branched, septate, subhyaline, smooth,  $1.5-2.5 \ \mu m$  wide. Setae absent. Conidiophores as on the natural substratum,  $(100-)127-176 \ \mu m$ long, 5.5–6.5(–7.5)  $\mu m$  wide in the middle and 13–16  $\mu m$  wide at the base, tapering to 5–6(–6.5)  $\mu$ m. Phialides with the proliferating part 5–14  $\mu$ m long and 5–6(–6.5)  $\mu m$  wide, with 10–12 proliferations. Conidia 19–22(–24)  $\times$  6–7  $\mu m$ . In 2-month old slant cultures the conidia became mature, the two middle cells gradually turning brown and the two end cells remaining hyaline or at least paler, mature conidia often sticking to the middle and lower part of the conidiophore.

Material examined. 1) Type material. Italy. Cansiglio (Treviso), on decorticated wood of Fagus sylvatica, Oct. 1874, P. A. Saccardo (PAD – holotype of Melanomma macrosporum). – USA, Georgia, Darien, on wood of Quercus sp., Ravenel 2420, (K 59126 – holotype of Sphaeria decastyla; NY – isotype).

2) Additional material. France, Central Pyrenees, Bagnères de Luchon, Lys valley, on decayed wood of *Fagus sylvatica*, 13 July 1997, M. R. (Herb. M. R. 985/97, 988/97). – USA, New Jersey, Newfield, on bark of *Magnolia* sp., 10 July 1887 (NY); New Jersey, Gloucester County, Newfield, 22 Jan. 1878 (NY); ibid., on a bark of *Magnolia* sp., 29 June 1882 (NY); Louisiana, on a decayed log of oak, 2 Jan. 1886, A. B. Langlois (NY).

Cultures. CBS 101313 (Herb. M. R. 985/97); CBS 101314 (Herb. M. R. 988/97).

Descriptions and illustrations. Saccardo (1875: 73; 1878: 449; 1883: 210, 221); Cooke (1878: 52); Ellis and Everhart (1882: 155); Berlese (1894: 100, Tab. 94, Fig. 1).

Habitat. Saprobe on decayed wood of deciduous trees.

Known hosts. Fagus sylvatica, Magnolia sp., Quercus sp.

Known distribution. Europe: France, Italy; North America: USA (New Jersey, Lousiana).

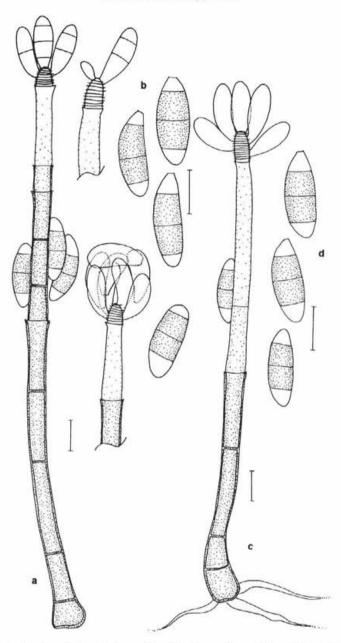


Fig. 9a–d. Chaetosphaeria decastyla. – a. Conidiophore with conidia and sporogenous conidiophore apices, from nature. – b. Conidia, from nature. – c. Conidiophore with conidia, from PCA culture. – a. d from Herb. M. R. 985/97; c, d from CBS 101313. – Scale bar: a–d = 10  $\mu$ m.

Nomenclatural note. The name Chaetosphaeria decastyla is based on Sphaeria decastyla Cooke 1878, even though the name Melanomma macrosporum Sacc. 1875 would have priority. If the epithet 'macrosporum' were transferred to Chaetosphaeria this would result in a homonymous combination (Art. 53.1, Greuter et al. 1994) with Chaetosphaeria macrospora (Kawamura) Hara [Hara, J. Pl. Prot. Tokyo 16(2): 16, 1930] that is based on Miyoshiella macrospora Kawamura [Kawamura, J. Coll. Sci. Imp. Univ. Tokyo 23(2): 295, 1929]. Miyshiella macrospora does not belong to Miyoshiella Kawamura, a member of the Trichosphaeriaceae G. Winter (Réblová 1998c). Miyshiella macrospora is a pyrenomycetous ascomycete forming black spots on bamboo culms having transversely 7-septate, blackish brown ascospores and unitunicate asci. Therefore, the second available name is chosen and the new combination Chaetosphaeria decastyla is proposed.

Note. The holotype and isotype and other collections of Sphaeria decastyla, including those cited by Ellis and Everhart (1882), are preserved in K and NY. They possess perithecia accompanied by conidiophores of the Cacumisporium capitulatum anamorph. Ascospores of North American collections, except for one specimen (USA, New Jersey, Gloucester County, Newfield, 22 Jan. 1878), differ in shape and size from European material that is represented by Saccardo's type material of Melanomma macrosporum and two recent collections made in France. Specimens from North America have rather fusiform and shorter ascospores (Fig. 8)  $[(28-)30-34.5(-36.5) \times 3-4 \ \mu\text{m}]$  than those of the European material, in which the ascospores (Fig. 7) are cylindrical to cylindrical-fusiform and longer  $[(31.5-)35.5-42(-46) \times 3-4 \ \mu\text{m}]$ . The specimen collected in USA, New Jersey, Gloucester County, Newfield, has ascospores much like those of the European collections, cylindrical, measuring  $(27-) 30.5-39(-42) \times 3-4 \ \mu\text{m}$ .

Ellis and Everhart (1882) described perithecia sparsely clothed with obtuse, septate setae. These setae (50–115 × 3–4.5  $\mu$ m) were found only on several perithecia of the holotype of *Sphaeria decastyla* and other material (USA, New Jersey, Newfield, on bark of *Magnolia* sp., 10 July 1887, NY). The majority of perithecia were glabrous and bore remnants of conidiophores. The obtuse setae were not found in European material. Ellis and Everhart (1882) noted that the substratum surface in both collections was clothed with hairs similar to those growing on the perithecia. Our revision revealed that the so-called hairs are conidiophores of the *Cacumisporium capitulatum* anamorph.

Ellis and Everhart (1882) cited in a synonymy of Acanthostigma decastylum three names, viz. Sphaeria cariosa Cooke et Ellis, Sphaeria atriella Cooke et Ellis and Lasiosphaeria subvelutina Ellis et Everhart. Sphaeria cariosa (Holotype: USA, New Jersey, Newfield, on bark of decayed oak, 15 Feb. 1877, J. B. Ellis 2789, K 59125, NY – isotype) is a species of Chaetosphaeria, known as Chaet. ovoidea (Fr.) O. Constant. et al.; the Menispora glauca Pers. anamorph is abundantly present on the type. Sphaeria atriella Cooke et Ellis (Isotype: USA, New Jersey,

Newfield, Gloucester County, on rotten wood of Acer sp., Dec 1876, NY) was combined by Barr (1993) under Chaetosphaeria, as Chaet. atriella (Cooke et Ellis) M. E. Barr. We do not believe that this is a true Chaetosphaeria species, for it differs in several characters that would characterize a Chaetosphaeria. The perithecial wall is leathery, three-layered, ca. 35–48  $\mu$ m thick, composed of nonopaque, thick-walled cells; paraphyses were not present and the asci were enclosed in a hyaline matrix that could be formed of dissolved paraphyses. No anamorph was associated. The wall of perithecium of Chaetosphaeria is different; it is thiner, ca. 15–25  $\mu$ m, comprising thin-walled, opaque brick-like cells or cells that form a network, paraphyses are always persistent and do not dissolve. Lasiosphaeria subvelutina is another possible synonym of Chaetosphaeria decastyla, but the type could not be examined (not available in NY).

The type material of *Melanomma macrosporum* contained several mature perithecia and conidiophores of the *Cacumisporium capitulatum* anamorph arising sparsely from the substratum. Mature bicolorous conidia were found attached to the substratum and to the perithecial walls.

The conidiogenesis of *Cac. capitulatum* was described in detail by Goos (1969). The conidia have a flat basal scar and are formed successively from multiple growing points on the apex of the conidiogenous cell. The conidiogenous cells proliferate conspicuously 5–14  $\mu$ m above the shallow collarette. A similarly proliferating apex of the conidiogenous cells is also found in the *Cylindrotrichum* anamorph of *Chaetosphaeria acutata*, which differs by smaller, hyaline, 2-celled conidia. The measurements of conidia of *Cac. capitulatum* are given from the original culture obtained immediately after isolation from the ascospores. Conidia are generally larger in vitro than on material from nature. After several transfers, *Cac. capitulatum* produces much smaller conidia in vitro [(14–)16–22(–24) x 5–7  $\mu$ m] than in nature.

The ascospores of *Chaetosphaeria decastyla* are somewhat similar in shape to those of *Chaet. crustacea. Chaet. decastyla* can be clearly distinguished from *Chaet. crustacea* by the exclusive occurrence on wood of angiosperms, longer ascospores and asci, and the *Cac. capitulatum* anamorph.

5. Chaetosphaeria fennica (P. Karst.) Réblová et W. Gams, comb. nov.

Figs. 10, 11.

- Easiosphaeria britzelmayri Sacc. subsp. fennica P. Karst., Rev. Mycol. Toulouse 9: 160, 1887. – basionym.
- $\equiv$  Acanthostigma fennicum (P. Karst.) Berlese, Icon. Fung. 1: 102, 1894.
- ≡ Zignoëlla abietis Höhnel var. fennica (P. Karst.) Höhnel, Sitzungsber. K. Akad. Wiss., Wien, Math.-naturw. Kl., Abt. 1, 118: 332, 1909.

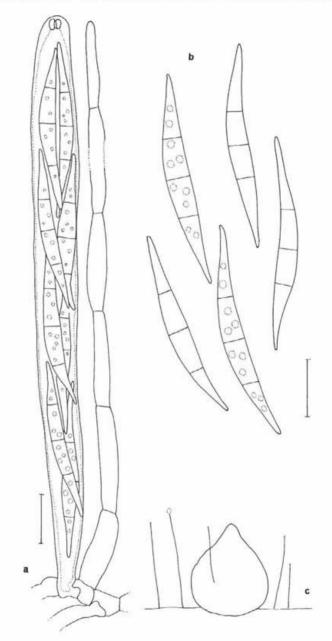


Fig. 10a-c. Chaetosphaeria fennica. – a. Ascus with ascospores and a paraphysis. – b. Ascospores. – c. Habit sketch of perithecium and conidiophores. – a-c from H 929. – Scale bars: a, b = 10  $\mu$ m.

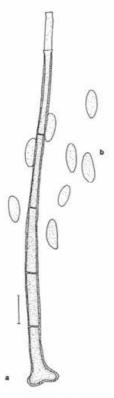


Fig. 11a, b. Chaetosphaeria fennica. – a. Conidiophore, from nature. – b. Conidia, from nature. – a, b from H 929. – Scale bar: a,  $b = 10 \ \mu m$ .

Anamorph. Chloridium-like (described here). Fig. 11a, b.

Teleomorph. Perithecia solitary, scattered, subglobose, 150–200  $\mu$ m high and 200–220  $\mu$ m diam, papillate, papilla perforated by a rounded pore in old perithecia, ostiolate, dark brown to black, glabrous or covered with conidiophores of the anamorph, smooth. Perithecial wall brittle, 22–26  $\mu$ m thick, consisting of two layers; an outer layer of thin-walled, dark brown, opaque, polyhedral to brick-like cells, and an inner layer of thin-walled, compressed, hyaline cells. Ostiolar canal periphysate. Paraphyses copious, branching, anastomosing, septate, 4–5  $\mu$ m wide in the lower part, tapering to 2.5  $\mu$ m, protruding slightly beyond the tips of the asci. Asci long-cylindrical, (126–)133–152(–168) × (8.5–)9–10.5  $\mu$ m, narrowly rounded at the tip, with a J-, refractive apical annulus, 2.5  $\mu$ m wide and 1–1.5  $\mu$ m deep. Ascospores elongate-fusiform and tapering strongly towards the ends immediately after the middle septum, (34.5–)36.5–42(–43) × (3.5–)4(–4.5)  $\mu$ m, at maturity 3septate, not constricted at the septa, smooth-walled, hyaline, 2-seriate, overlapping partially in the upper part of the ascus.

Anamorph. Remnants of conidiophores with broken apices were observed growing copiously from the substratum surface and occasionally from the outer wall of the perithecia. Conidiophores macronematous, mononematous, erect, solitary, unbranched, septate, opaque, paler to subhyaline towards the top, 250–270  $\mu$ m long, 5.5–6.5  $\mu$ m wide above the base and 20–22  $\mu$ m wide at the base. Once, a conidiophore with an inconspicuously proliferated apex ending in a hyaline collarette 3  $\mu$ m wide and 2  $\mu$ m deep was observed. A lot of conidia were seen attached to the conidiophore. Conidia mid-brown, ellipsoidal, straight or inequilateral, slightly truncate at the base, 8.5–9.5(–10) × 3–4.5  $\mu$ m.

The teleomorph-anamorph connection could not yet be proved by culture studies and is only suggested by the joint occurrence of perithecia and conidiophores.

Material examined. Type material. Finland, Mustiala Myllyperä, on decayed bark of *Betula* sp., on old stromata of *Eutypa* sp., 28 July 1887, K. Starbäck, herb. P. Karsten 929 (H – holotype of *Chaetosphaeria fennica*).

Descriptions and illustrations. Karsten (1887: 160); Berlese (1894: 102, Pl. 119, Fig. 3); Saccardo (1891: 852); Podlahová (1974: 149, Tab. 42).

Habitat. Saprobe on stromata of Eutypa sp.

Known host. Eutypa sp. on Betula sp.

Known distribution. Europe: Finland, known only from the type locality.

Note. Podlahová (1974) revised the type material and her observations agree well with ours. Although Karsten (1887) and Berlese (1894) described and illustrated the perithecia as sparsely setose, Podlahová (1974) found them being glabrous. The present revision of the type material showed that the presumed setae were conidiophores that are usually quite firm but only visible as remnants in old herbarium material.

Lasiosphaeria britzelmayri Sacc. (Saccardo 1883) differs from Chaetosphaeria fennica in having shorter asci (90–100 × 9–10  $\mu$ m) and shorter (28–35 × 3.5–4  $\mu$ m), 6–11-celled, cylindrical-fusiform ascospores. Winter (1887) considered this fungus to have bitunicate asci and classified it as Trematosphaeria paradoxa G. Winter.

On the basis of ascospore anatomy, *Chaetosphaeria fennica* is closest to *Chaet. acutata*. The ascospores of the latter, though also long-fusiform, are much less tapering towards the ends and are rather shorter. The *Cylindrotrichum* anamorph of *Chaet. acutata* with hyaline and 1-celled conidia at maturity is entirely different from the *Chloridium*-like anamorph associated with *Chaet. fennica*.

### 6. Chaetosphaeria fusiformis W. Gams et Hol.-Jech., Mycotaxon 13: 257, 1981.

Figs. 12, 13.

E Chaetosphaeria fusispora W. Gams et Hol.-Jech., Stud. Mycol. 13: 45, 1976. (illegitimate, Art. 53.1.) [non Chaetosphaeria fusispora (Kawamura) Hino,

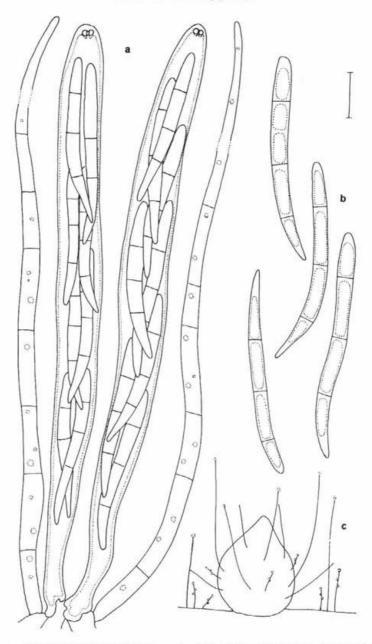


Fig. 12a–c. Chaetosphaeria fusiformis. – a. Asci with ascospores and paraphyses. – b. Ascospores. – c. Habit sketch of the perithecium and conidiophores. – a–c from Herb. M. R. 862/96. – Scale bar: a, b = 10  $\mu$ m.

Bull. Miyazaki Coll. Agr. For. 4: 191, 1932, nec Chaetosphaeria fusispora P. Larsen, Dansk Bot. Ark. 14(7): 7, 1952.]

Anamorph. Chloridium cylindrosporum W. Gams et Hol.-Jech., Stud. Mycol. 13: 46, 1976. Figs. 13a–c.

≡ Chaetopsis cylindrospora (W. Gams et Hol.-Jech.) DiCosmo et al., Mycologia 75: 962, 1983.

Teleomorph. Perithecia superficial, densely gregarious or in small groups, globose, 170–250  $\mu$ m high and 160–240  $\mu$ m diam, papillate, ostiolate, black, glistening, glabrous, covered with conidiophores of the anamorph, slightly rugose. Perithecial wall brittle, lateral wall 18–28  $\mu$ m thick, consisting of two layers; an outer layer of dark brown, thin-walled, opaque, brick-like cells, and an inner layer of thin-walled, subhyaline, compressed cells. Ostiolar canal periphysate. Paraphyses copious, septate at 10–21  $\mu$ m intervals, slightly constricted at the septa, branching, 3–5  $\mu$ m wide in the lower part, tapering to 2–2.5  $\mu$ m, rounded at the top, protruding beyond the tips of the asci. Asci cylindrical-clavate, (89–)99–126(–146) × 8.5–10.5(–11.5)  $\mu$ m , shortly stipitate, ascal apex with a J-, refractive, 2.5–3  $\mu$ m wide and 1–1.5  $\mu$ m deep apical annulus that is situated ca. 1  $\mu$ m below the apex. Ascospores fusiform, tapering at one end and rounded at the other, rarely tapering to both ends, (34.5–)39–53.5(–62) × 2.5–3(–4)  $\mu$ m, 3-septate, not constricted at the septa, smooth-walled, hyaline, 2–3-seriate, overlapping in the ascus.

Anamorph. Colonies hairy, dark, conidiophores arising from the substratum or covering the perithecia. Setae absent. Conidiophores macronematous, mononematous, solitary, erect, unbranched, forming two layers. Conidiophores of the lower layer cylindrical, 33–55  $\mu$ m tall and 3.5–4  $\mu$ m wide in the middle, tapering to 1.5–2 (–2.5)  $\mu$ m below the collarette, straight or slightly flexuous, 0–2-septate, pale brown, sympodially proliferating. Conidiophores of the upper layer cylindrical, (88–)116–230  $\mu$ m tall and 6–7  $\mu$ m wide above the base, tapering to 2.0–2.5  $\mu$ m below the collarette, straight, 6–10-septate, thick-walled, brown to dark brown, paler upwards, usually with 1 percurrent proliferation, ending in a monophialide. Phialides terminal, integrated or intercalary, polyblastic, with 1 apical or 1–3 lateral phialidic openings arising from the sympodial proliferation. Collarettes hyaline, 4–5  $\mu$ m wide and 2–3  $\mu$ m deep. Conidia phialidic, cylindrical, centrally slightly constricted, tapering towards the ends, truncate at the base, 10.5–13.5 × 3.5–5  $\mu$ m, aseptate when young, later forming 2–3 additional septa, smooth-walled, hyaline, in dry, irregular or star-like heads.

Characteristics in culture. Colonies on PCA slow-growing, reaching 3 mm diam in 10 days at 25 °C in darkness; when grown for another 10 days at 25 °C in

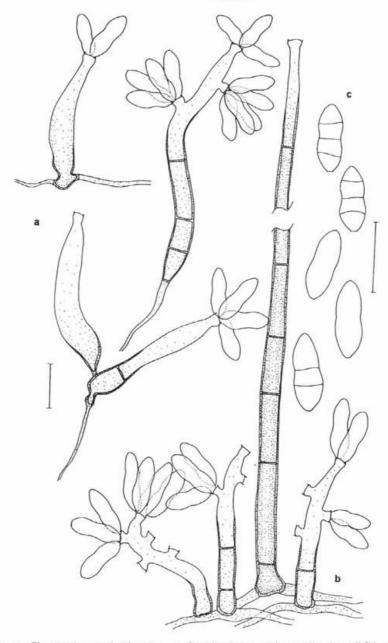


Fig. 13a–c. Chaetosphaeria fusiformis. – a. Conidiophores with conidia, from PCA culture. – b. Taller and shorter conidiophores with conidia forming two distinct layers, from nature. – c. Conidia, from nature. – a from CBS 101430; b, c from Herb. M. R. 862/96. – Scale bar:  $a-c = 10 \ \mu m$ .

## Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

cool white fluorescent light reaching 4–5 mm diam; velvety, mid-brown, zonate, aerial mycelium developed, margins fimbriate; conidial production copious in 20 days, conidiophores scattered, conidial masses globose, hyaline; reverse pale brown. CMA, OA: 4–5 mm diam, velvety, zonate, aerial mycelium developed, conidial production copious in the centre in 10 days; CMA: ochraceous to pale brown; OA: mid-brown; MEA: 2–2.5 mm diam, felty, not zonate, mid-brown, aerial mycelium scant, no conidiation in 20 days. Mycelium superficial or immersed; hyphae branched, septate, subhyaline, smooth-walled, 1.5–2.5  $\mu$ m wide. Setae absent. Conidiophores as on the natural substratum, conidiophores of the lower layer (24–)30–59(–65)  $\mu$ m tall and 5–6(–7)  $\mu$ m wide above the base, tapering to 1.5–2.5  $\mu$ m; conidiophores of the upper layer up to 136  $\mu$ m tall and 6–7  $\mu$ m wide above the base, tapering to 2.5–3  $\mu$ m. Phialides with collarettes 4–5  $\mu$ m wide and 2–3  $\mu$ m deep. Conidia 11.5–13.5(–17) × 4–5  $\mu$ m.

An upper layer of longer conidiophores was only seen in 4-months old slant cultures on PCA preserved at 18 °C under cool white fluorescent light.

Material examined. 1) Type material. Czech Republic. Moravia, Hrubý Jeseník Mts., on slopes of Mt. Mravenečník near Loučná nad Desnou, on decayed branch of *Abies alba*, 3 Aug. 1971, V. Holubová-Jechová (PRM 794008 – holotype of *Chaetosphaeria fusispora* ( $\equiv$  *Chaetosphaeria fusiformis*).

2) Additional material. Czech Republic. Southern Bohemia, Sumava Mts., glacial cirque of the lake Černé jezero near Železná Ruda, on the inner surface of bark of *Abies alba*, 23 Oct. 1996, M. R. (Herb. M. R. 862/97, 863/97, 866/96, 867/96, 868/96, 874/96, 889/96); ibid., 27 Aug. 1997, M. R. (Herb. M. R. 1042/97, 1048/97, 1049/97); ibid., glacial cirque of the lake Čertovo jezero near Železná Ruda, on bark of *Abies alba*, 22 Oct. 1996, M. R. (Herb. M. R. 864/96). Moravia, Hrubý Jeseník Mts., on slopes of Mt. Mravenečník near Loučná nad Desnou, on decorticated wood of *Abies alba*, 3 Aug. 1971, V. Holubová-Jechová (PRM 794111). – Ukraine. Eastern Carpathian Mts., Kvasi near Rachiv, on the left bank of the Tisa River, on the inner side of bark on a stump of *Abies alba*, 26 June 1997, M. R. (Herb. M. R. 949/97, 958/97).

Cultures. CBS 101429 (Herb. M.R. 1048/97), CBS 101430 (Herb. M.R. 1049/97).

Descriptions and illustrations. Gams and Holubová-Jechová (1976: 45; Figs. 21a, b, 22).

Habitat. Saprobe on decayed wood and inner surface of bark of gymnosperms. Known host. *Abies alba*.

Known distribution. Europe: Czech Republic, Ukraine.

Note. Although the conidia of *Chloridium cylindrosporum* were described as non-septate (Gams and Holubová-Jechová 1976), 3-septate conidia were seldom observed on recent material from nature. The 3-septate conidia can easily escape our

attention because they are usually attached to the lower part of the conidiophores, the surface of the perithecia or the substratum. The presence of conidial septation, although the septa are formed much later, would argue for placement of the *Chloridium cylindrosporum* anamorph of *Chaet. fusiformis* in *Cylindrotrichum*. Because no applicable and final generic concept of *Cylindrotrichum* is available and the relationships between *Cylindrotrichum* and *Chloridium* require further analysis, we refrain at the moment from making this combination. However, *Chaetosphaeria fusiformis* is considered another member of a group of related species of *Chaetosphaeria* with *Cylindrotrichum* anamorphs.

*Chaetosphaeria fusiformis* is highly host-specific and occurs seldom, exclusively on decayed wood and bark of coniferous trees. All recent collections were made in regions with natural stands of *Abies alba*.

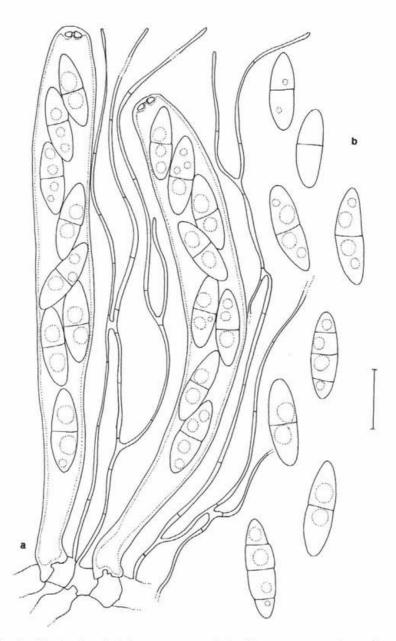
Of Chaetosphaeria species with Cylindrotrichum anamorphs, Chaet. abietis and Chaet. crustacea also occur exclusively on coniferous wood and bark. Chaet. fusiformis differs from these two species in having longer, at maturity 3-septate and slightly asymmetrical ascospores that taper towards one end and are rounded at the other end, and the Chloridium cylindrosporum anamorph. The ascospores of Chaet. abietis and Chaet. crustacea are symmetrical and cylindrical-fusiform, 3-septate in the former and 3-5-septate in the latter.

This is the first report of successful cultivating of *Chaet. fusiformis*. The anamorph-teleomorph connection, previously suggested on the basis of the regular joint occurrence, is herewith confirmed.

# 7. Chaetosphaeria tulasneorum Réblová et W. Gams, sp. nov. Figs. 14–16.

Perithecia superficialia, ad basim modice immersa, solitaria vel pauca aggregata, subglobosa vel conica, deorsum applanata, 190–220  $\mu$ m alta, 185–210  $\mu$ m diam, minute papillata, ostiolata, fusca, glabra, modice verrucosa. Canalis ostiolaris periphysatus. Paries perithecii fragilis, ad latus et apicem scleroticus, deorsum attenuatus; paries lateralis 26–39  $\mu$ m crassus, bistratosus: stratum exterius e cellulis tenuitunicatis, fuscis, lateriformibus compositum, interius e cellulis tenuitunicatis, compressis, subhyalinis. Paraphyses copiosae, filiformes, raro septatae, ad septa haud constrictae, ramosae, anastomosantes, reticulum formantes, hyalinae, 12  $\mu$ m latae in parte inferiore, ultra ascorum apices protrudentes. Asci cylindrici vel clavati, 93.5–115 × 8.5–10.5(–12.5)  $\mu$ m, sursum modice truncati, breviter stipitati, apex anulo I-, refringente, 2  $\mu$ m lato et 0.5–1  $\mu$ m alto praeditus, 8-spori. Ascosporae fusiformes, 15.5–20(–21) × 4–5(–6)  $\mu$ m, plerumque bicellulares, sero 2 alteris septis divisa, haud vel paene constrictae in medio, leves, hyalinae, 1–2-seriatae in asco.

Anamorphosis Cylindrotrichum oligospermum (Corda) Bonord.



Réblová M. and Gams W.: Teleomorph-anamorph connections in Ascomycetes

Fig. 14a, b. Chaetosphaeria tulas neorum. – a. Asci with as cospores and paraphyses. – b. Ascospores. – a, b from PRM 842978. – Scale bar: a, b = 10  $\mu m.$ 

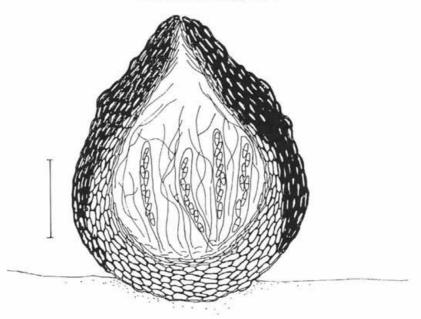


Fig. 15. Chaetosphaeria tulasneorum. – Median, longitudinal section of perithecium, note the sclerotisation of the lateral perithecial wall. – From PRM 842978. – Scale bar: 50  $\mu$ m.

Holotypus. Bohemia meridio-occidentalis, montes Javornická hornatina, Strašín apud Sušice, ad lignum putridum *Sambuci nigrae*, 21 Oct. 1997, leg. M. Svrček (PRM 842978).

Anamorph. Cylindrotrichum oligospermum (Corda) Bonord., Handb. allg. Mykol. p. 88, 1851. Fig. 16a-e.

- $\equiv$  Menispora oligosperma Corda, Icon. Fung. 2: 12, 1838.
- = Acrothecium deliculatum Berk. et Broome, Ann. Mag. nat. Hist., Ser. 3, 15: 402, 1865.
  - ≡ Cordana deliculata (Berk. et Broome) O. Kuntze, Rev. Gen. Pl. 2: 850, 1891.

Etymology. Honouring the Tulasne brothers, L.-R. Tulasne and C. Tulasne, who described the genus *Chaetosphaeria*.

Teleomorph. Perithecia superficial, at the base slightly immersed, solitary or in small groups, subglobose to conical, base flattened, 190–220  $\mu$ m high and 185–210  $\mu$ m diam, minutely papillate, ostiolate, dark brown, glabrous, slightly verrucose. Perithecial wall brittle, the whole wall heavily sclerotised in the upper part, sclerotisation disappearing towards the bottom and absent at the base. 34

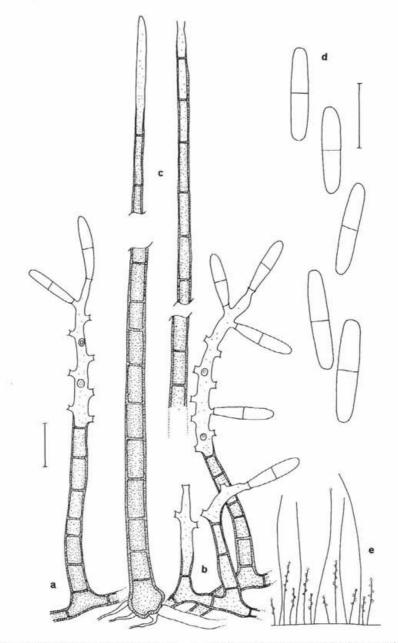


Fig. 16a–e. Chaetosphaeria tulasneorum. – a–c. Taller and shorter conidiophores with conidia forming two distinct layers and setae, from PCA culture. – d. Conidia, from PCA culture. – e. Habit sketch of conidiophores and setae. – a–e. from CBS 101319, PRM 842978. – Scale bar: a–d = 10  $\mu$ m.

Lateral wall 26–39  $\mu$ m thick, consisting of two layers; an outer layer of thinwalled, dark brown, brick-like cells, and an inner layer of thin-walled, compressed, subhyaline cells. Ostiolar canal periphysate. Paraphyses copious, filiform, sparsely septate, not constricted at the septa, forming a branching network, hyaline,  $1-2 \ \mu$ m wide, protruding beyond the tips of the asci. Asci cylindrical to clavate,  $93.5-115 \times 8.5-10.5(-12.5) \ \mu$ m, slightly truncate at the top, shortly stipitate, ascal apex with a J-, refractive,  $2 \ \mu$ m wide and  $0.5-1 \ \mu$ m deep apical annulus, 8-spored. Ascospores fusiform,  $15.5-20(-21) \times 4-5(-6) \ \mu$ m, mostly 2-celled, with a delayed formation of the 2 additional septa, not constricted or slightly constricted at the median septum, smooth, hyaline, 1-2-seriate, partially overlapping in the ascus.

Anamorph. Remnants of shorter and longer conidiophores and setae with broken apices were found growing from the perithecia and from the substratum surface on a decorticated branch of *Sambucus nigra*.

Characteristics in culture. Colonies on PCA slow-growing, reaching 27 mm diam in 10 days at 25 °C in darkness, when grown for another 10 days at 25 °C in cool white fluorescent light reaching 15-16 mm diam; velvety, greyish-yellowish in the centre due to crowded conidiophores, yellowish at the margins, margins fimbriate, thin, aerial mycelium scant, conidiophores scattered, copious conidial production in 10 days, conidial masses whitish; reverse yellow-greyish. CMA, OA: 21-25 mm diam; MEA: 18 mm diam; CMA, OA, MEA: velvety, aerial mycelium scanty, conidial production copious in 10 days; CMA, OA: yellowish to pale brown in the centre due to crowded conidiophores, whitish sterile zone at the margins; MEA: whitish to pale brown in the centre due to crowded conidiophores, withish sterile zone at the margins. Mycelium superficial or immersed; hyphae branched, septate, subhyaline, smooth, 2–3  $\mu$ m wide. Setae straight, cylindrical, 180–560  $\mu$ m tall and 3–4.5  $\mu$ m wide in the middle, tapering to 1.5–2  $\mu$ m, 5–23-septate, dark brown, paler towards the apex, narrowly rounded at the top or some developing into a monophialide. Conidiophores macronematous, mononematous, solitary, erect, unbranched, arising from both aerial and substrate mycelium, forming two layers. Conidiophores of the lower layer 33–66(–104)  $\mu m$  tall and 3.5–4.5  $\mu m$ wide in the middle, tapering to 1.5–2.5  $\mu$ m below the collarette, cylindrical, straight or slightly flexuous, 0-4-septate, subhyaline to pale brown, sympodially proliferating. Conidiophores of the upper layer 180–280  $\mu m$  tall and 3–4.5  $\mu m$ wide in the middle, paler towards the apex, tapering to  $1.5-2 \ \mu m$  below the collarette, cylindrical, straight or slightly flexuous, 10-12-septate, mid-brown to dark brown, less commonly sympodially proliferating at the top. Phialides integrated, terminal or intercalary, with 6-11 lateral phialidic openings arising from a sympodial elongation, fertile apices  $23-47(-57) \times 3.5-4(-4.5) \mu m$ . Collarettes hyaline, disappearing early, 2–2.5  $\mu$ m wide and 2–3  $\mu$ m deep. Conidia phialidic,

cylindrical to clavate,  $(9-)11-14.5(-17) \times (2-) 2.5-3(-4) \mu m$ , 1-septate, not constricted, rounded at the tip, tapering and truncate at the base, smooth, hyaline.

Material examined. Type material. Czech Republic. South-western Bohemia, Javornická hornatina Mts., Strašín near Sušice, on wood of a dead branch of Sambucus nigra, 21 Oct. 1997, M. Svrček (PRM 842978 – holotype of Chaetosphaeria tulasneorum).

Culture. CBS 101319 (Herb. M. R. 1164/97).

Habitat. Saprobe on decayed wood of a shrub.

Known host. Sambucus nigra.

Known distribution. Europe: Czech Republic; known only from the type locality.

Note. Shape and septation of the ascospores of *Chaetosphaeria tulasneorum* resemble those of *Chaet. innumera* Berk. et Broome ex Tul. et C. Tul., *Chaet. pulviscula* (Currey) C. Booth and *Chaet. ovoidea* (Fr.) O. Constant. et al., but the ascospores are longer than those of the first and shorter than those of the two latter species. *Chaet. tulasneorum* is also distinct in the *Cylindrotrichum oligospermum* anamorph and the occurrence on decorticated wood of an angiosperm shrub. The lateral perithecial wall has a very characteristic sclerotisation that was not observed in any other *Chaetosphaeria* species.

We are not aware of any report of a *Cylindrotrichum oligospermum* anamorph isolated from ascospores of any pyrenomycetous ascomycete. The connection to the teleomorph is reported here for the first time by a culture study. The conidiophores obtained in the living culture formed two layers of taller and shorter conidiophores accompanied by much longer setae. The upper layer of conidiophores was observed neither on the natural substratum nor in culture by Gams and Holubová-Jechová (1976). In all other respects our isolate of *Cyl. oligospermum* agrees well with the observations of Gams and Holubová-Jechová (1976) and Barron (1968).

The size of setae, shorter conidiophores, phialides, collarettes and conidia are given from the original culture of Cyl. oligospermum obtained immediately after the isolation from ascospores. The description of taller conidiophores that have 8–11 lateral phialidic openings is given from an older culture after several transfers. In the original culture fertile apices with well-developed lateral phialidic openings were hardly seen.

### DISCUSSION

The division of *Cylindrotrichum* into five genera, viz. *Chaetopsis*, *Dictyo-chaeta*, *Kylindria*, *Xenokylindria* and *Uncigera*, and their delimitation proposed by DiCosmo et al. (1983), seems to be rather schematic, apparently disregarding

natural units. Chaetopsis grisea (Ehrenb.) Sacc. has holoblastic conidiogenesis and a predominantly lateral branching pattern of the conidiophores unlike Cylindrotrichum oligospermum and is regarded as representing a distinct genus. Uncigera cordae Sacc. cannot be separated from Cylindrodendrum album Bonord. (W. Gams, unpublished), as redescribed by Buffin and Hennebert (1984). The remaining genera are very closely related indeed. Cylindrotrichum is closest to Chloridium Link: Fr., from which it is merely separated by conidial septation and shape. Conidia in Cylindrotrichum are 1- to multi-septate, usually cylindrical to long-ellipsoidal, straight or slightly curved and truncate at the base. Conidia of *Chloridium* are non-septate, generally shortly ellipsoidal, rarely fusiform to clavate (Chloridium codinaeoides Pirozynski) or shortly catenate to dacryoid [Chloridium clavaeforme (Preuss) W. Gams et Hol.-Jech.] or reniform (Chloridium reniforme Matsushima). But longer ellipsoidal conidia have also been described, e.g. in Chloridium matsushimae W. Gams et Hol.-Jech. (1976: 29). Thus the anamorphs of Chaetosphaeria crustacea and Chaet. fusiformis might be classified in either genus. Cylindrotrichum accommodates species with both sympodial and percurrent proliferation of the conidiophores (Cabello and Arambarri 1988); Cyl. oligospermum (Corda) Bonord., the type species of the genus, has sympodially proliferating conidiophores with numerous lateral phialidic openings, the conidiophores are accompanied by sterile setae and the conidiogenesis is multiple from different loci within a collarette. The presence of percurrent besides sympodial conidiophore proliferation would argue for distinguishing two sections within Cylindrotrichum, comparable with the situation in Chloridium. Chloridium was divided into three sections (Gams and Holubová-Jechová 1976), viz. species with sympodial proliferation of conidiophores with single conidiogenous loci (sect. *Psilobotrys*), and percurrent proliferation of conidiophores with multiple (sect. Chloridium) or single conidiogenous loci (sect. Gongromeriza). Species of both Cylindrotrichum and Chloridium are currently associated with Chaetosphaeria and Melanopsammella Höhnel species (Booth 1957, 1958; Gams and Holubová-Jechová 1976; Réblová et al. 1999).

The conidiogenesis of the Cacumisporium capitulatum anamorph of Chaetosphaeria decastyla is identical to that of the Cylindrotrichum anamorph of Chaet. acutata, the Cylindrotrichum anamorph of Chaet. crustacea, and the Chloridium virescens (sect. Chloridium) anamorph of Melanopsammella vermicularioides (Sacc. et Roum.) Réblová et al. (Réblová et al. 1999). However, the conidiophores of Cac. capitulatum proliferate percurrently and the conidia are 3-septate and bicolorous. The conidia are formed successively from multiple conidiogenous loci on the sporogenous apex of the percurrently proliferating conidiogenous cell. The conidia mature after being liberated and usually remain attached to the conidiophores separately or in groups of 2–3. In the Cylindrotrichum anamorph of Chaet. acutata and the Cac. capitulatum anamorph of Chaet. decastyla the

sporogenous apex of the conidiogenous cell has a similar function and structure as in *Xenokylindria* if a generic separation were required (DiCosmo et al. 1983).

Nine genera have been described for anamorphs of *Chaetosphaeria* (Réblová et al. 1999). In the present paper *Cacumisporium* is introduced as another anamorph genus. The anamorph genera of *Chaetosphaeria* are more or less closely related to each other and require further analysis. Until this has been done, we retain the Saccardoan treatment for them. In Saccardo's (1886) scheme conidial septation was a major criterion and thus we retain *Chloridium*, *Cylindrotrichum*, and *Cacumisporium* for the time being as separate though very closely related taxa.

## ACKNOWLEDGMENTS

We wish to thank the Directors and Curators of the H, NY, PAD and PRM herbaria for loaning material. We are grateful to Dr. Mirko Svrček for providing the Latin diagnosis of *Chaetosphaeria acutata*. This project was partly supported by the Grant Agency of Charles University in Prague (26266/96), Grant "Centres of Biological Diversity in the Šumava Mts. Biosphere Reserve" from the World Bank supported by GEF (Global Environment Facility) and by the Grant "Monitoring of lignicolous Ascomycetes in glacial cirques of the lakes in Šumava Mts." of Agency for nature conservation and landscape protection.

# References

- BARR M. E. (1990): Prodromus to non-lichenized, pyrenomycetous members of class Hymenoascomycetes. – Mycotaxon 39: 43–184.
- BARR M. E. (1993): Redisposition of some taxa described by J. B. Ellis. Mycotaxon 46: 45–76. BARR M. E. and CRANE J. L. (1979): Another conidial state for a species of Chaetosphaeria. –
- Can. J. Bot. 57: 835–837.
- BARRON G. L. (1968): The Genera of Hyphomycetes from soil. Williams and Wilkins Co., Baltimore, 364 pp.
- BERLESE A. N. (1894): Icones fungorum omnium hucusque cognitorum. Vol. 1. (Reprint Biblioth. Mycol. 16A: 1–243.).

BOOTH C. (1957): Studies on pyrenomycetes I. and II. - Mycol. Pap. 68: 1-27.

BOOTH C. (1958): The genera Chaetosphaeria and Thaxteria in Britain. - Naturalist 1958: 83-90.

BUFFIN N. and HENNEBERT G. L. (1984): Cylindrodendrum album Bonorden, a pleoanamorphic semiaquatic hyphomycete. – Mycotaxon 19: 323–341.

CABELLO M. and ARAMBARRI A. (1988): Considerations about the validity of the genus Cylindrotrichum Bonorden. – Mycotaxon 31: 435–438.

CLEMENTS F. E. and SHEAR C. L. (1931): The genera of fungi. - New York, 496 pp.

CONSTANTINESCU O., HOLM K. and HOLM L. (1995): Teleomorph-anamorph connections in Ascomycetes: the anamorphs of three species of Chaetosphaeria. – Mycol. Res. 99: 585–592. COOKE M. C. (1878): Ravenel's American Fungi. – Grevillea 7: 32–54.

- DICOSMO F., BERCH S. and KENDRICK W. B. (1983): Cylindrotrichum, Chaetopsis, and two new genera of hyphomycetes, Kylindria and Xenokylindria. – Mycologia 75: 949–973.
- ELLIS J. B. and EVERHART B. M. (1892): North American Pyrenomycetes. Newfield, 793 pp. (Reprinted 1966).
- ERIKSSON O. E. and HAWKSWORTH D. L. (1993): Outline of the Ascomycetes 1993. Syst. Ascom. 12: 51–257.

GAMS W. and HOLUBOVÁ-JECHOVÁ V. (1976): Chloridium and some other dematiaceous hyphomycetes growing on decaying wood. – Stud. Mycol. 13: 1–99.

GAMS W. and HOLUBOVÁ-JECHOVÁ V. (1981): Chloridium and some other dematiaceous hyphomycetes growing on decaying wood. Corrections and additions. – Mycotaxon 13: 257–258.

GOOS R. D. (1969): Conidium ontogeny in Cacumisporium capitulatum. – Mycologia 61: 52–56. GREUTER W. et al. (1994): International Code of Botanical Nomenclature. – Regnum Vegetabile 131: [1–18], 1–389.

HAWKSWORTH D. L. and MINTER D. W. (1980): New and interesting microfungi from the 1978 Exeter Foray. – Trans. Brit. Mycol. Soc. 74: 567–577.

HÖHNEL F. VON (1909): Fragmente zur Mykologie 7. – Sitzungsber. K. Akad. Wiss. Wien, Math.naturw. Kl. 118: 275–452.

HOLMGREN P. K., HOLMGREN N. H. and BARNETT L. C. (1990): Index herbariorum. Part I: The herbaria of the world. Eighth edition. – Regnum Vegetabile vol. 120, p.(1)-(10), 1–693, New York.

HOLUBOVÁ-JECHOVÁ V. (1973): Lignicolous hyphomycetes from Czechoslovakia 4. Menispora. – Folia Geobot. Phytotax. 8: 317–336.

HOLUBOVÁ-JECHOVÁ V. (1982): New or interesting phialidic hyphomycetes from Cuba. – Mycotaxon 15: 277–292.

HOLUBOVÁ-JECHOVÁ V. (1984): Lignicolous hyphomycetes from Czechoslovakia 7. Chalara, Exochalara, Fusichalara and Dictyochaeta. – Folia Geobot. Phytotax. 19: 387–438.

HOLUBOVÁ-JECHOVÁ V. (1990): Problems in the taxonomy of the dematiaceous hyphomycetes. – Stud. Mycol. 32: 41–48.

HUGHES S. J. (1951): Stachylidium, Gonytrichum, Mesobotrys, Chaetopsis and Chaetopsella. – Trans. Brit. Mycol. Soc. 34: 551–576.

HUGHES S. J. and KENDRICK W. B. (1968): New Zealand fungi 12. Menispora, Codinaea, Menisporopsis. – New Zealand J. Bot. 6: 323–375.

KARSTEN P. A. (1873): Mycologia fennica, pars secunda. – Bidr. Känn. Finl. Nat. Folk 23: 1–252. KARSTEN P. A. (1887): Ascomycetes novi fennici. – Rev. Mycol. Toulouse 9: 159–177.

KARSTENT, A. (1997). The gradient invite internet. – Rev. Mycol. Toulouse 5, 135-111.

KENDRICK W. B. (1980): The generic concept in Hyphomycetes – a reappraisal. – Mycotaxon 11: 339–364.

MORGAN-JONES G. (1977): Notes on Hyphomycetes XVIII. Chaetoblastophorum ingramii gen. et sp. nov. and Cylindrotrichum oblongisporum. – Mycotaxon 5: 484–490.

MORGAN-JONES G. (1980): Notes on Hyphomycetes XXXIX. Cylindrotrichum curvatum sp. nov. – Mycotaxon 12: 249–252.

MÜLLER E. and ARX J. A. VON. (1962): Die Gattungen der didymosporen Pyrenomyceten. – Beitr. Krypt.-Fl. Schweiz 11(2): 1–922.

PODLAHOVÁ R. (1974): Lignicolous members of the Lasiosphaeriaceae (Fuck.) Nannf. I, II. – 320 pp., ms. in Czech, [PhD thesis; deposited in: Fac. Nat. Sci., Charles University, Prague].

RÉBLOVÁ M. (1998a): A new Chaetosphaeria with a Dictyochaeta anamorph. – Czech Mycol. 50: 151–159.

RÉBLOVÁ M. (1998b): Revision of three Melanomma species described by L. Fuckel. – Czech Mycol. 50: 161–179.

RÉBLOVÁ M. (1998c): Studies in Chaetosphaeria sensu lato III. The genera Umbrinosphaeria gen. nov. and Miyoshiella with Sporidesmium anamorphs. – Mycotaxon 71 (in press).

RÉBLOVÁ M., BARR M. E. and SAMUELS G. J. (1999): Chaetosphaeriaceae, a new family for Chaetosphaeria and its relatives. – Sydowia (in press).

Rенм H. (1907): Ascomycetes novi. - Ann. Mycol. 5: 516-546.

SACCARDO P. A. (1875): Fungi Veneti novi vel critici. Serie 3. - Hedwigia 14: 68-76.

SACCARDO P. A. (1878): Fungi Veneti novi vel critici. Serie 3. - Michelia 1: 446-452.

SACCARDO P. A. (1883): Pyrenomyceteae. Sylloge Fungorum omnium hucusque cognitorum 2: 1–813, Padova.

SACCARDO P. A. (1886): Hyphomyceteae. – Sylloge Fungorum omnium hucusque cognitorum 4: 1–807, Padova.

SACCARDO P. A. (1891): Supplementum universale. – Sylloge Fungorum omnium hucusque cognitorum 9: 1–1141, Padova.

TULASNE L. R. and TULASNE C. (1863): Selecta Fungorum Carpologia, Vol. 2. – Paris, 319 pp.

WINTER G. (1887): Die Pilze, Ascomyceten: Gymnoasceen und Pyrenomyceten. – In: Rabenhorst's Kryptogamen-Flora Deutschl., Österr. und der Schweiz, 2nd ed., Band 2, 1(2): 1–928, Leipzig.