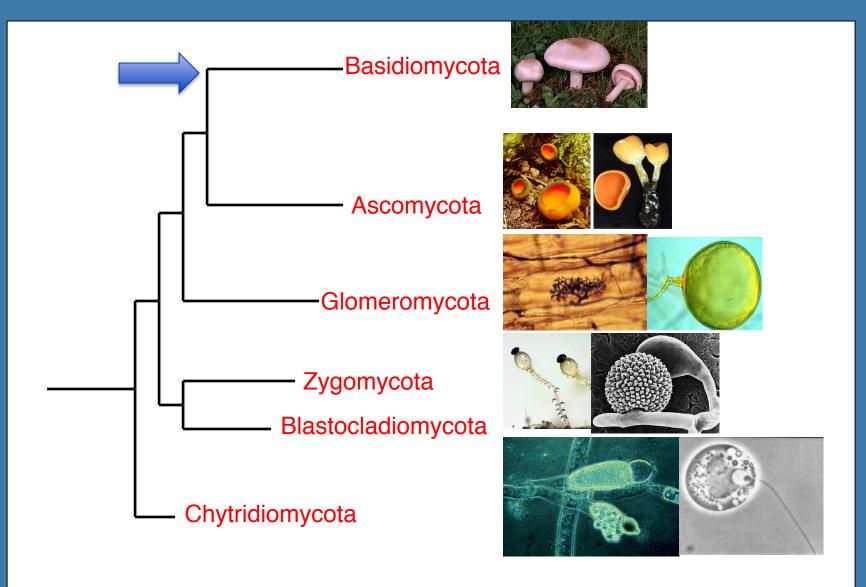
3. Basidiomycota: Life cycle and mating systems



Phyla of Fungi



Basidiomycota ~31,500 species

Diverse ecologies:

Saprobes/decomposers/wood decay Plant parasites/pathogens Animal and human pathogens Insect parasites/symbionts Mycorrhizal plant symbionts Nematode trappers/predators **Diversity in structures** Yeast (single cell) and filamentous forms Mushrooms, clubs, corals, stinkhorns, puffballs Perennial conks (polypores) Resupinate crusts (on decaying wood) Jelly fungi Truffles (hypogeous) Plant galls

Phylum Basidiomycota

- <u>b</u>asidiomycetes— common name for this phylum and no longer carries any formal taxonomic significance
- <u>Basidiomycota</u> (phylum) characterized by having meiospores called <u>basidiospores</u> that are formed on specialized cells, <u>basidia</u> (sing. basidium), the site of meiosis
- Three subphyla recognized <u>Current name (subphyla)</u> Names used in Webster (classes)
 - Agaricomycotina
 - Ustilaginomycotina
 - Pucciniomycotina

(Homobasidio<u>mycetes</u>) (Ustilagino<u>mycetes</u>

(Urediniomycetes)

Terminations of taxon names indicate rank

3 Major Clades - Subphyla - of the Basidiomycota

Agaricomycotina mushrooms, polypores, jelly fungi, corals, chanterelles, crusts, puffballs, stinkhorns

Ustilaginomycotina smuts, *Exobasidium*, Malassezia

Pucciniomycotina rusts, *Septobasidium*







Three Subphyla of Basidiomycota

Agaricomycotina (Hombasidiomycetes)

Agaricales, Boletales, Russulales, Cantharellales - mushrooms Gomphales, Phallales - coral, club fungi, stinkhorns Hymenochaetales, Polyporales - polypores Auriculariales, Dacrymycetales & Tremellales - jelly fungi Geastrales – earth stars

Ustilaginomycotina (Ustilaginomycetes)

Ustilaginomycetes (smuts) - plant pathogens Exobasidiomycetes –*Exobasidium* - plant pathogens *Malassezia* – mammalian skin yeasts

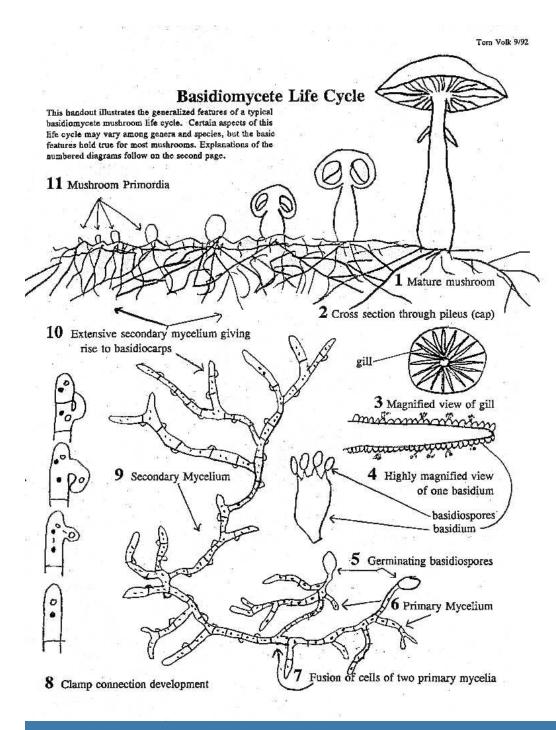
Pucciniomycotina (Urediniomycetes)

Pucciniomycetes, Pucciniales (=Uredinales, rusts) - plant pathogens Septobasidiales (crusts) - insect parasites Microbotryomycetes – plant pathogens









Major stages of Agaricomycete life cycle

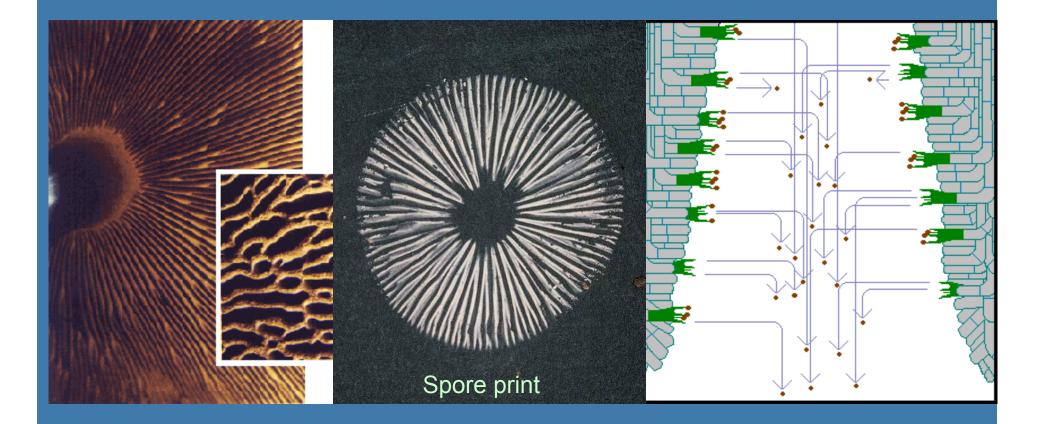
- 1) <u>Sporocarp</u>
- 2) Gilled Hymenophore
- 3) Hymenium
- 4) Basidium
- 5) Basidiospore
- 6) Monokaryon
- 7) Fusion of monokaryons
- 8) Clamp connections
- 9) Dikaryon
- 10) Mycelium
- 11) Sporocarp primordia

1. Sporocarp

Sporocarp – general term for macroscopic fungal reproductive structure, "fruiting body" Basidiocarp – term for sporocarp of the basidiomycetes Basidiome(ata) - formal term used to designate basidiomycete sporocarp(s) Hymenophore – a basidiocarp with a <u>hymenium</u> (layer of fertile cells)



2. Gilled hymenophore



Gills increase surface area for spore production up to 20 times that of a flat surface

1° 2° 3° lamellae

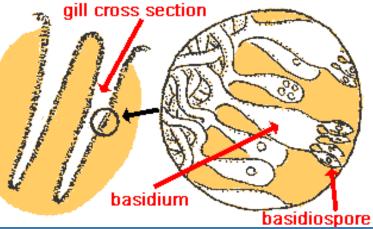


Spacing of lamellae in gilled sporocarps varies by species May have ony primary lamellae, or additional levels, secondary, tertiary

3. Hymenium

The tissue layer that contains the reproductive cells (basidia) and sterile supporting cells (cystidia). Gill



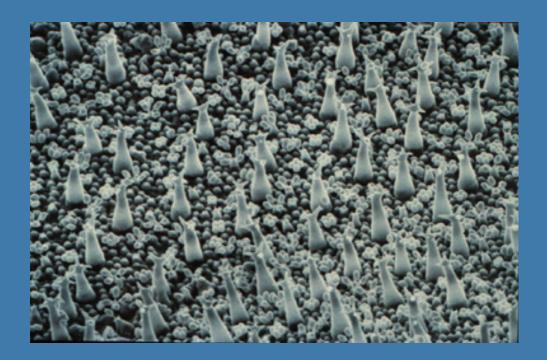


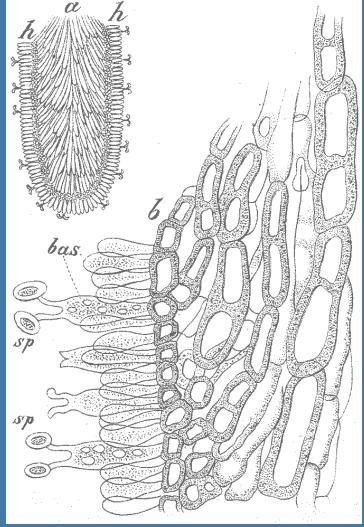
Pore



Hymenium

- Term for the organized tissue layer that produces basidiospores
- Basidia
- Basidioles
 - Cells resembling basidia that have not produced basidiospores
- Cystidia
 - Larger than other hymenial elements
 - Variety of shapes—taxonomically useful
 - Function not known





Hymenium/hymenophore

- Polypores—hymenium lining pores or tubes
- Chanterelles—hymenium on gill-like folds
- Toothed fungi—hymenium on small spines
- Coral fungi—erect basidiocarps, hymenium covering surface
- Corticioid fungi (resupinate)—smooth or wrinkled hymenium
- Agarics—hymenium on "gills" or lamellae
- "Jelly fungi" smooth or wrinkled hymenium



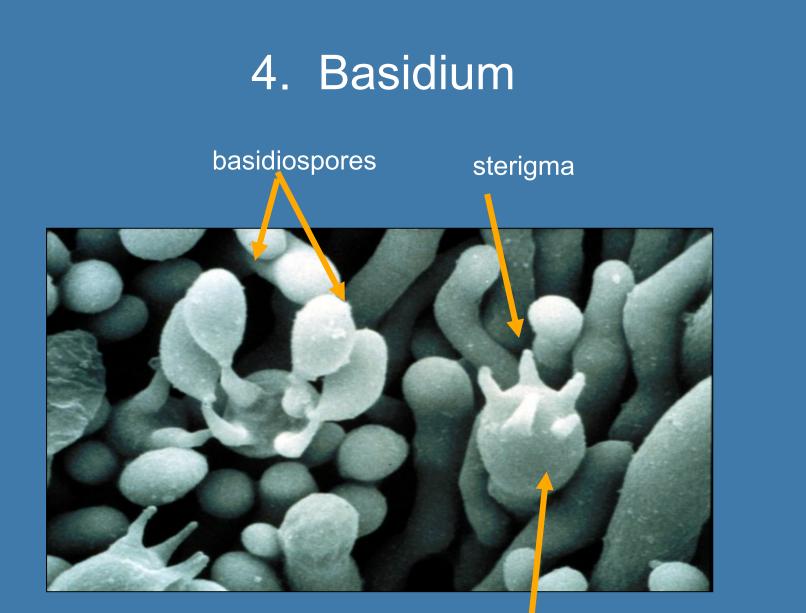










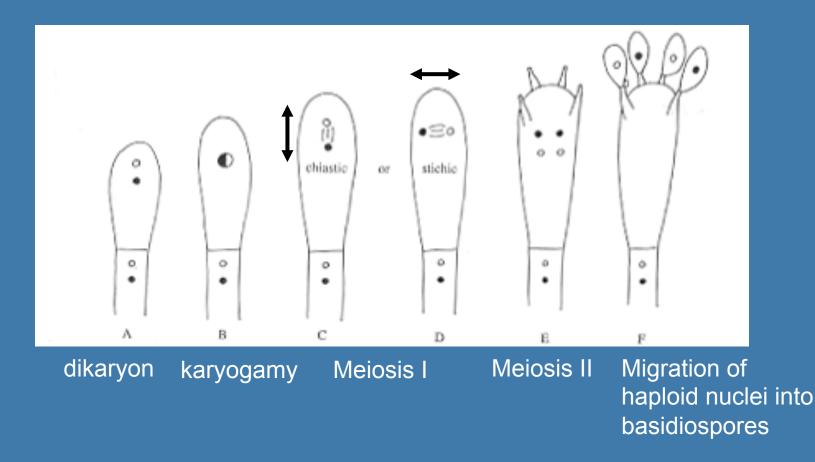


basidium

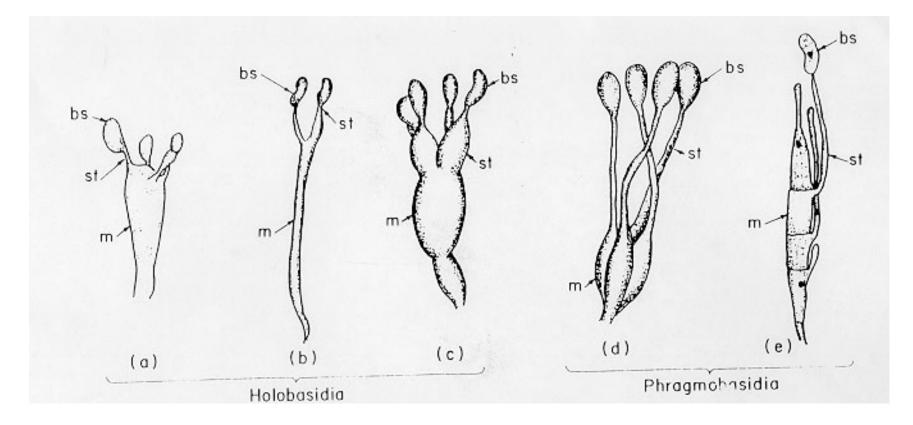
4. Basidium

• • •

- Defining structure of the Basidiomycota
- Site of meiosis; products are basidiospores
- Usually located in specialized regions or tissues (hymenium)

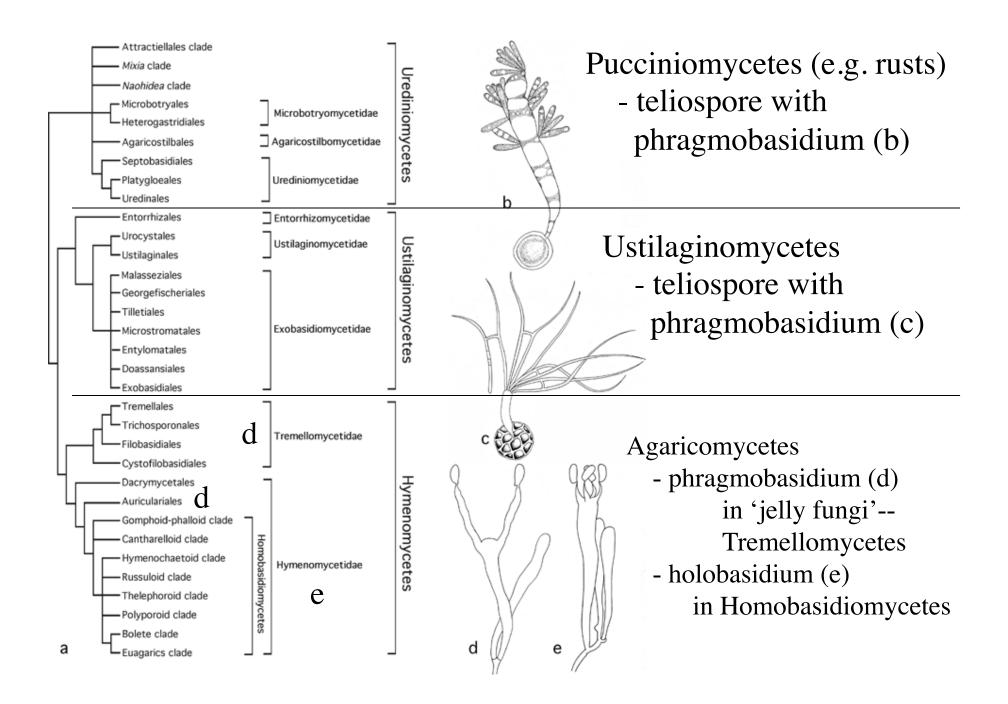


Types of basidia



Holobasidia: basidium not divided by septa

Phragmobasidia: Basidium divided by one or more septa, transverse or cruciate





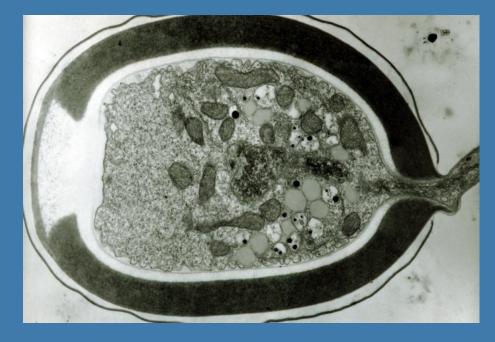


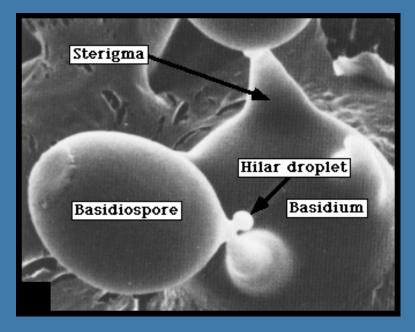
Note that basidiospores are attached asymmetrically

5. Basidiospore

Dissemination: "surface tension catapult"

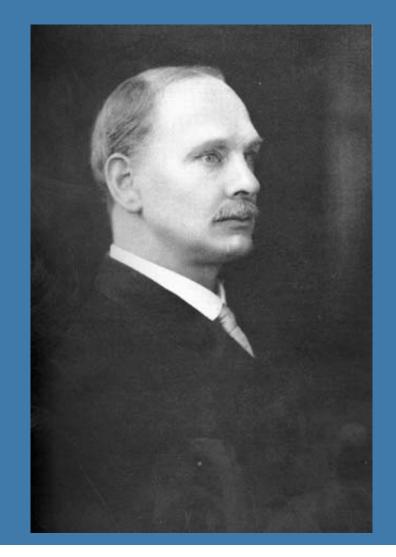
- 1. hygroscopic substances secreted near hilar appendix, cause water condensation, hydrophobic wall region keeps drops apart
- 2. fusion of <u>Buller's drop</u> and adaxial drop causes rapid displacement of the center of mass of the spore
- 3. this exerts a force that is opposed by the sterigma
- 4. the spore is projected away from the sterigma





Buller's Drop

A. H. Reginald Buller



Researches on Fungi 1909

Studied spore liberation, genetics and pretty much everything about fungi, and wrote scientific limericks

There was a young lady named Bright Whose speed was far faster than light; She set out one day, In a relative way And returned on the previous night.

To her friends said the Bright one in chatter, "I have learned something new about matter: My speed was so great, Much increased was my weight, Yet I failed to become any fatter!"

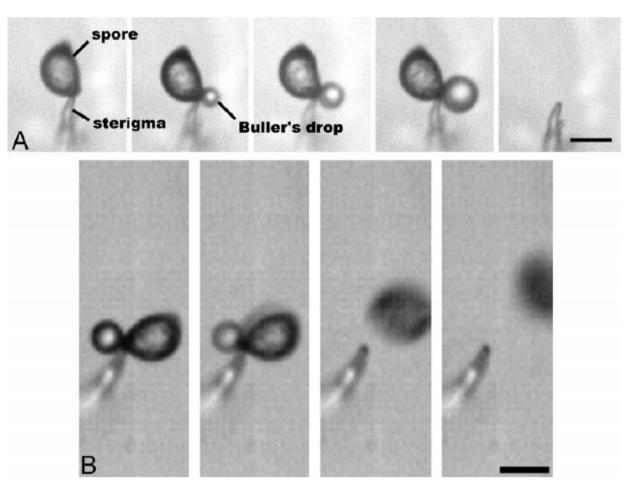
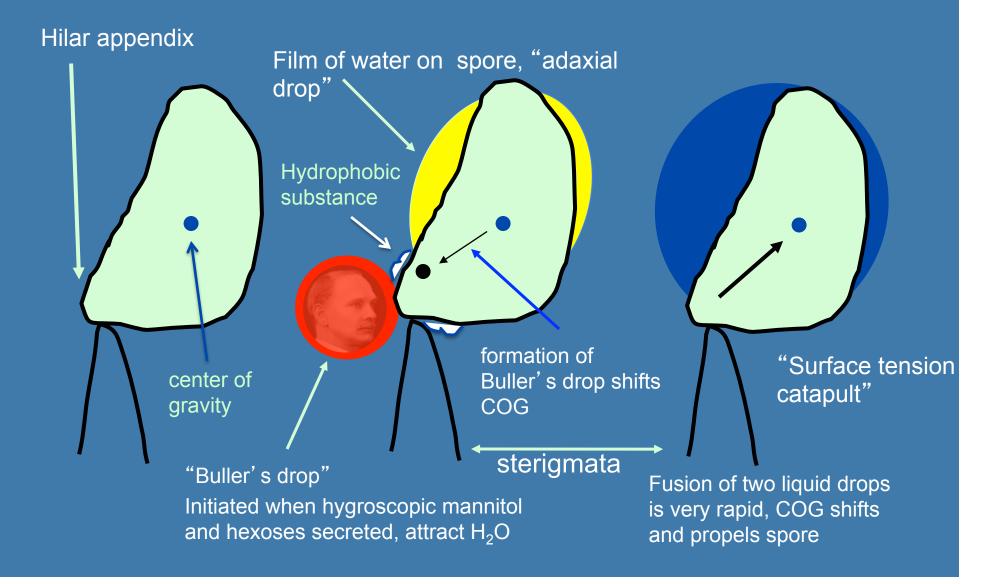
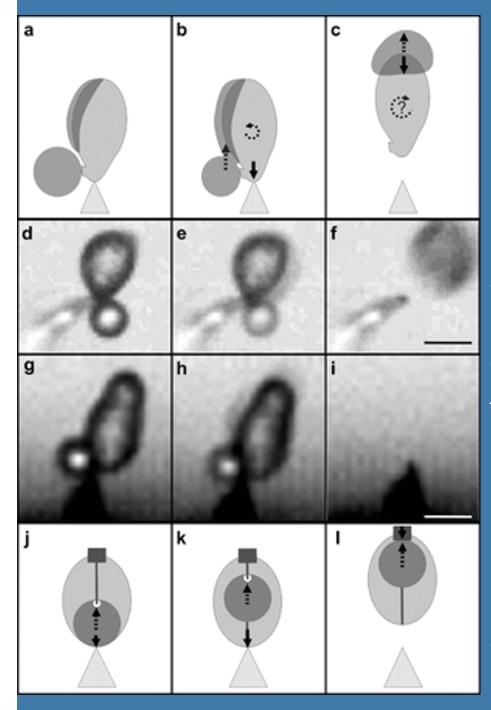


FIG. 1. a. Mechanism of ballistospore discharge in *I. perplexans* as captured using conventional still photomicroscopy. Successive images separated by 10 s show the growth of Buller's drop and simultaneous disappearance of drop and spore from sterigma in final frame (scale bar 10 μ m). Photographs courtesy of John Webster. b, Images of Buller's drop and the ballistospore using ultra high speed video. Successive images separated by 10 μ s (scale bar 10 μ m).

5. Basidiospore

Mechanism of propulsion: Buller's drop & surface tension





a: Buller's drop contains energy in the form of surface tension forces.

b: When Buller's drop touches the fluid on the side of the spore (the adaxial drop), the merging fluid flows rapidly toward the distal end of the spore

c: The collapsed drop and spore launch from the sterigma.

d–e: An oblique view of the spore and Buller's drop of *I. perplexans* (10 μ s between each frame, scale bar 10 μ m).

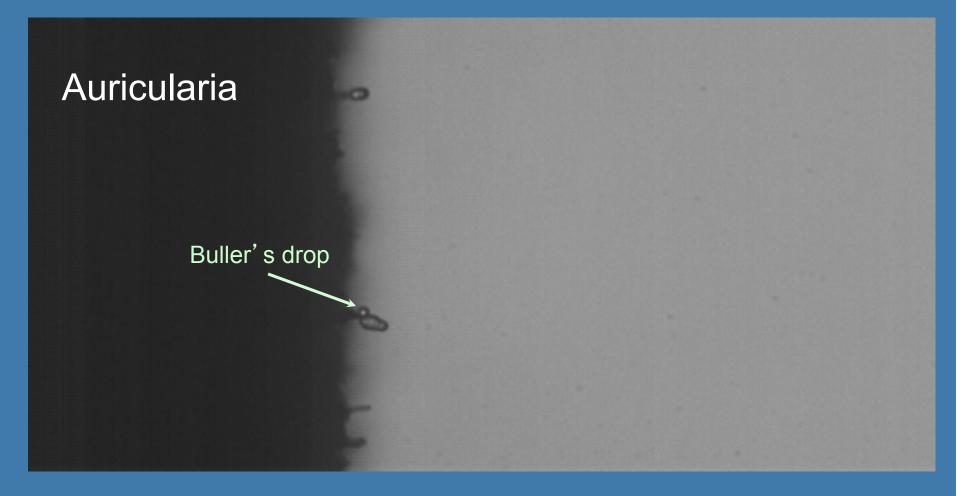
g–I; A lateral view of *A. auricularia* during ballistospore launch (50 μ s between each frame, scale bar 10 μ m).

j: the drop begins to collapse and surface tension energy is released.

I: surface tension forces cause the drop to stop abruptly and prevent the fluid from flying off of the spore. The abrupt deceleration delivers a directed and substantial force to the spore, and causes the spore and drop to eject away from the sterigma. The increasing drop momentum during its directed distal movement is critical for directing the spore away from the sterigma, and for delivering sufficient force for spore release.

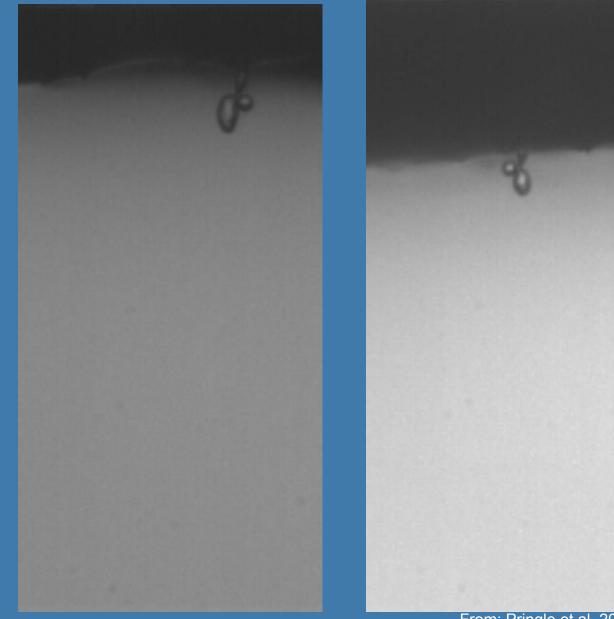
From: Pringle et al. 2005 Mycologia 97: 866-871

Basidiospore Release



From: Pringle et al. 2005 Mycologia 97: 866-871

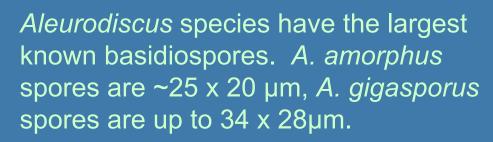
High speed video micrographs of basidiospore release



From: Pringle et al. 2005 Mycologia 97: 866-871

How far and how fast can basidiospores travel?





Volume of a *A. gigasporus* spore would be $1.4 \times 10^{-14} \text{ m}^3$

At a density of 1.2 x 10³ kg per m³, these spores would have a mass of about 17ng

Xerula radicata has the largest known spores of a gilled mushroom, $17 \times 14 \mu m$.

see Fischer et al 2010, Fungal Biology 114:669-675



How far and how fast can basidiospores travel?



Tectella patellaris has the smallest known basidiospores of a gilled basidiomycete, 3.7×0.7 . *Hyphodontia latitans* has $3.5 - 5 \times 0.5 - 0.8 \mu m$ spores.

At a volume of 0.5 x 10^{-19} m³ these spores would have a mass of 0.6 pg (~10,000 times less than *A. gigasporus*).

How far and how fast can basidiospores travel?

Fischer et al. (Fungal Biology 114:669-675) used high speed video to measure the velocity of basidiospore discharge by large and small-spored species. To estimate distance traveled, drag force, air viscosity, spore radius and mass are the parameters needed.

Species	Spore size (µm)	Spore volume	Buller' s drop radius (µm)	Est. spore mass (g)	velocity (m per sec)	range (mm), spore lengths
Aleurodiscus gigasporus	34 x 28	1.4 x 10 ⁻¹⁴	8.3	1.9 x 10⁻ ⁸	0.53	1.83, 54
Xerula radicata	17 x 14	1.6 x 10 ⁻¹⁵	3.7	2.2 x 10 ⁻⁹	0.70	0.58, 35
Trametes versicolor	5 x 2	1.4 x 10 ⁻¹⁸	0.6	8.5 x 10 ⁻¹²	0.68	0.01, 3
Hyphodontia latitans	4 x 0.6	1.4 x 10 ⁻¹⁹	0.3	6.2 x 10 ⁻¹³	1.05	0.004, 1

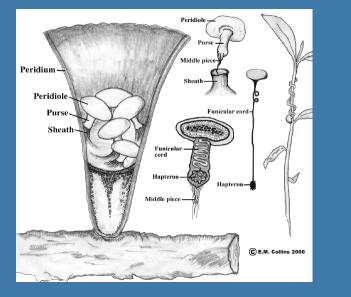
"Hymenomycetes" vs. "Gasteromycetes"

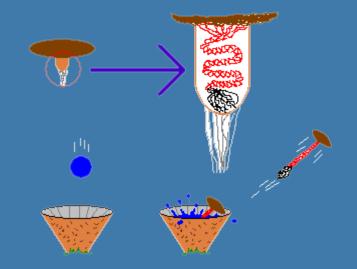
As used informally, both are polyphyletic.

Hymenomycetes: general term for taxa having <u>holobasidia</u> and a definite hymenium that remains intact during sporulation Hymenium exposed, gills, pores etc Basidiospores actively discharged ballistospores

Gasteromycetes: No distinct hymenium present at time of basidiospore release Closed basidiocarps Basidiospores passively discharged Included in Class Agaricomycotina

5. Basidiospore Dissemination: Passive (rain drop) Bird's nest fungus







Common taxa

Crucibulum

- Cup-shaped, dull white peridioles, funiculus



Expulsion of peridiole by a water drop

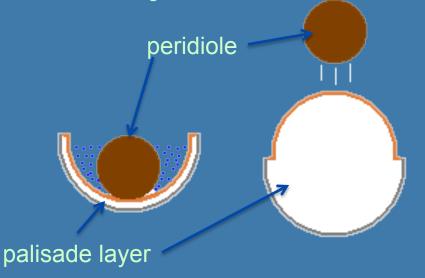


5. Basidiospore

Dissemination: Active Cannon ball fungus, *Sphaerobolus*

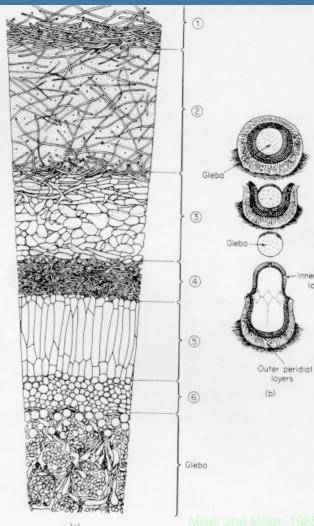


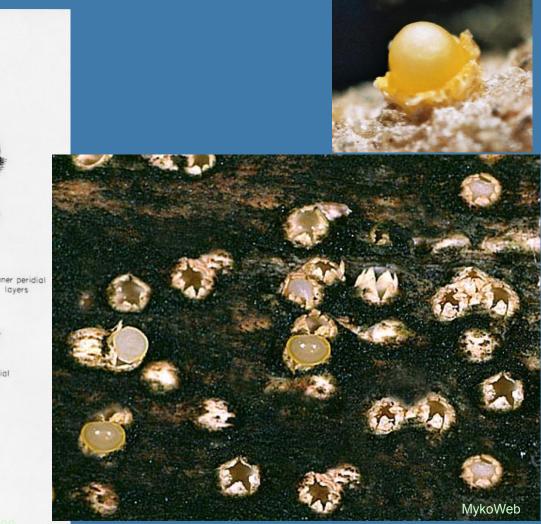
Conversion of glycogen to glucose in palisade layer cells causes increase in osmotic pressure. Movement of H_2O into the palisade results in increased turgor pressure and explosive expulsion of peridiole as the inner cup truns inside out. Peridiole is sticky and adheres to vegetation.

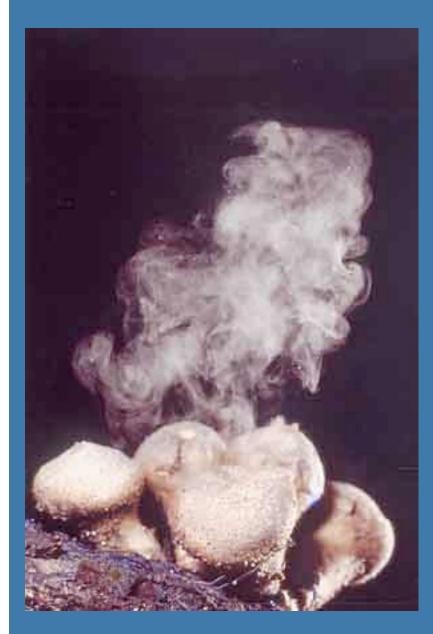


Sphaerobolus

 Cannon-ball fungus; one peridiole, forcibly discharged by evagination of endoperidium







Other gasteromycetes such as *Lycoperdon* passively disperse basidiospores when a water drop strikes the thin persistent peridium

Mycelium

- Three types of mycelia occur in the basidiomycete life cycle:
 - Primary mycelium (monokaryon)
 - Germinated basidiospore
 - Secondary mycelium (dikaryon)
 - Anastomosis of compatible mating type monokaryons
 - Characterized by clamp connections in many taxa
 - Tertiary mycelium
 - organized, specialized tissues that make up the basidiocarp